At the high dose (1.2 µg), the glycoPEGylated rFSH had somewhat higher *in vivo* activity than the unPEGylated rFSH.

### G-CSF

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### 28. GlycoPEGylation of G-CSF produced in CHO cells

Preparation of Asialo-Granulocyte-Colony Stimulation Factor (G-CSF). G-CSF produced in CHO cells is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 5 mM CaCl $_2$  and concentrated to 500  $\mu L$  in a Centricon Plus 20 centrifugal filter. The solution is incubated with 300 mU/mL Neuraminidase II ( $\it Vibrio\ cholerae$ ) for 16 hours at 32 °C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel performed. The reaction mixture is then added to prewashed N-(p-aminophenyl)oxamic acid-agarose conjugate (800 μL/mL reaction volume) and the washed beads gently rotated for 24 hours at 4 °C. The mixture is centrifuged at 10,000 rpm and the supernatant was collected. The beads are washed 3 times with Tris-EDTA buffer, once with 0.4 mL Tris-EDTA buffer and once with 0.2 mL of the Tris-EDTA buffer and all supernatants are pooled. The supernatant is dialyzed at 4 °C against 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub> and then twice more against 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub>. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter and stored at -20 °C. The conditions for the IEF gel were run according to the procedures and reagents provided by Invitrogen. Samples of native and desialylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of G-CSF-(alpha2,3)-Sialyl-PEG. Desialylated G-CSF was dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of ST3Gal1 at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis

according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

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Preparation of G-CSF-(alpha2,8)-Sialyl-PEG. G-CSF produced in CHO cells, which contains an alpha2,3-sialylated O-linked glycan, is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN3, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of CST-II at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction has CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of G-CSF-(alpha2,6)-Sialyl-PEG. G-CSF, containing only O-linked GalNAc, is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of ST6GalNAcl or II at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction has CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

G-CSF produced in CHO cells was treated with Arthrobacter sialidase and was then purified by size exclusion on Superdex75 and was treated with ST3Gal1 or ST3 Gal2 and then with CMP-SA-PEG 20Kda. The resulting molecule was purified by ion exchange and

gel filtration and analysis by SDS PAGE demonstrated that the PEGylation was complete. This is the first demonstration of glycoPEGylation of an O-linked glycan.

#### Glucocerebrosidase

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29. Glucocerebrosidase-mannose-6-phosphate produced in CHO cells

This example sets forth the procedure to glycoconjugate mannose-6-phosphate to a peptide produced in CHO cells such as glucocerebrosidase.

Preparation of asialo-glucoceramidase. Glucocerebrosidase produced in CHO cells is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, and is incubated with 300 mU/mL sialidase-agarose conjugate for 16 hours at 32 °C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel and SDS-PAGE performed according to Invitrogen procedures. The mixture is centrifuged at 10,000 rpm and the supernatant is collected. The beads are washed 3 times with Tris-EDTA buffer, once with 0.4 mL Tris-EDTA buffer, and once with 0.2 mL of the Tris-EDTA buffer. All supernatants are pooled. The supernatant is dialyzed at 4 °C against 50 mM Tris-HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub> and then twice more against 50 mM Tris-HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub>. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Glucocerebrosidase-SA-linker-Mannose-6-phosphate (procedure 1). Asialo-glucocerebrosidasefrom above is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-linker-Man-6-phosphate and 0.1 U/mL of ST3Gal3 at 32°C for 2 days. To monitor the incorporation of sialic acid-linker-Man-6-phosphate, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas TSK-Gel-3000 analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using

SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Glucocerebrosidase-SA-linker-Mannose-6-phosphate (procedure 2). Glucocerebrosidase, produced in CHO but incompletely sialylated, is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-linker-Man-6-phosphate and 0.1 U/mL of ST3Gal3 at 32°C for 2 days. To monitor the incorporation of sialic acid-linker-Man-6-phosphate, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas TSK-Gel-3000 analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

## 30. Glucocerebrosidase-transferrin

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This example sets forth the procedures for the glycoconjugation of proteins, and in particular, transferrin is glycoconjugated to glucocerebrosidase. The GlcNAc-ASN structures are created on glucoceraminidase, and Transferrin-SA-Linker-Gal-UDP is conjugated to GNDF GlcNAc-ASN structures using galactosyltransferase.

Preparation of GlcNAc-glucocerebrosidase (Cerezyme<sup>TM</sup>). Cerezyme<sup>TM</sup> (glucocerebrosidase) produced in CHO cells is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, and is incubated with 300 mU/mL Endo-H-agarose conjugate for 16 hours at 32 °C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel and SDS-PAGE performed according to Invitrogen procedures. The mixture is centrifuged at 10,000 rpm and the supernatant is collected. The beads are washed 3 times with Tris-EDTA buffer, once with 0.4 mL Tris-EDTA buffer and once with 0.2 mL of the Tris-EDTA buffer and all supernatants are pooled. The supernatant is dialyzed at 4 °C against 50 mM Tris –HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub> and then twice

more against 50 mM Tris –HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub>. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Transferrin-SA-Linker-Gal-glucocerebrosidase. Transferrin-SA-Linker-Gal-UDP from above is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl<sub>2</sub>, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 2.5 mg/mL GlcNAc-glucocerebrosidaseand 0.1 U/mL of galactosyltransferase at 32°C for 2 days. To monitor the incorporation of glucocerebrosidase, the peptide is separated by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1) and the product detected by UV absorption. The reaction mixture is then purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

#### GM-CSF

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# 31. Generation and PEGylation of GlcNAc-ASN Structures: GM-CSF produced in Saccharomyces

This example sets forth the preparation of Tissue-type Activator with PEGylated GlcNAc-Asn structures.

Recombinant GM-CSF expressed in yeast is expected to contain 2 N-linked and 2 O-linked glycans. The N-linked glycans should be of the branched mannan type. This recombinant glycoprotein is treated with an endoglycosidase from the group consisting of endoglycosidase H, endoglycosidase-F1, endoglycosidase-F2, endoglycosidase-F3, endoglycosidase-M either alone or in combination with mannosidases I, II and III to generate GlcNAc nubs on the asparagine (Asn) residues on the peptide/protein backbone.

The GlcNAc-Asn structures on the peptide/protein backbone is then be modified with galactose or galactose-PEG using UDP-galactose or UDP-galactose-6-PEG, respectively, and a galactosyltransferase such as GalT1. In one case the galactose-PEG is the terminal residue.

In the second case the galactose is further modified with SA-PEG using a CMP-SA-PEG donor and a sialyltransferase such as ST3GalIII. In another embodiment the GlcNAc-Asn structures on the peptide/protein backbone can be galactosylated and sialylated as described above, and then further sialylated using CMP-SA-PEG and an  $\alpha 2.8$ -sialyltransferase such as the enzyme encoded by the *Campylobacter jejuni* cst-II gene.

### Herceptin<sup>TM</sup>

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## 32. Glycoconjugation of mithramycin to Herceptin™

This example sets forth the procedures to glycoconjugate a small molecule, such as mithramycin to Fc region glycans of an antibody molecule produced in mammalian cells. Here, the antibody Herceptin<sup>TM</sup> is used, but one of skill in the art will appreciate that the method can be used with many other antibodies.

Preparation of Herceptin<sup>TM</sup>-Gal-linker-mithramycin. Herceptin<sup>TM</sup> is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl<sub>2</sub>, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM UDP-galactose-linker-mithramycin and 0.1 U/mL of galactosyltransferase at 32°C for 2 days to introduce the mithramycin in the Fc region glycans. To monitor the incorporation of galactose, a small aliquot of the reaction has <sup>14</sup>C-galactose-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The fractions containing product are combined, concentrated, buffer exchanged and then freeze-dried. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

### Interferon α and Interferon β

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33. GlycoPEGylation of Proteins expressed in Mammalian or Insect Systems: EPO, Interferon  $\alpha$  and Interferon  $\beta$ 

This example sets forth the preparation of PEGylated peptides that are expressed in mammalian and insect systems.

Preparation of acceptor from mammalian expression systems. The peptides to be glycoPEGylated using CMP-sialic acid PEG need to have glycans terminating in galactose. Most peptides from mammalian expression systems will have terminal sialic acid that first needs to be removed.

Sialidase digestion. The peptide is desialylated using a sialidase. A typical procedure involves incubating a 1 mg/mL solution of the peptide in Tris-buffered saline, pH 7.2, with 5 mM CaCl<sub>2</sub> added, with 0.2 U/mL immobilized sialidase from *Vibrio cholera* (Calbiochem) at 32°C for 24 hours. Microbial growth can be halted either by sterile filtration or the inclusion of 0.02% sodium azide. The resin is then removed by centrifugation or filtration, and then washed to recover entrapped peptide. At this point, EDTA may be added to the solution to inhibit any sialidase that has leached from the resin.

Preparation from insect expression systems. EPO, interferon-alpha, and interferon-beta may also be expressed in non-mammalian systems such as yeast, plants, or insect cells. The peptides to be glycoPEGylated using CMP-sialic acid PEG need to have glycans terminating in galactose. The majority of the N-glycans on peptides expressed in insect cells, for example, are the trimannosyl core. These glycans are first built out to glycans terminating in galactose before they are acceptors for sialyltransferase.

Building acceptor glycans from trimannosyl core. Peptide (1 mg/mL) in Trisbuffered saline, pH 7.2, containing 5 mM MnCl<sub>2</sub>, 5 mM UDP-glcNAc, 0.05 U/mL GLCNACT II, is incubated at 32°C for 24 hours or until the reaction is substantially complete. Microbial growth can be halted either by sterile filtration or the inclusion of 0.02% sodium azide. After buffer exchange to remove UDP and other small molecules, UDP-galactose and MnCl<sub>2</sub> are each added to 5 mM, galactosyltransferase is added to 0.05 U/mL, and is incubated at 32°C for 24H or until the reaction is substantially complete. Microbial growth can be halted either by sterile filtration or the inclusion of 0.02% sodium azide. The peptides are then ready for glycoPEGylation.

Building O-linked glycans. A similar strategy may be employed for interferon alpha to produce enzymatically the desired O-glycan Gal-GalNAc. If necessary, GalNAc linked to serine or threonine can be added to the peptide using appropriate peptide GalNAc transferases (e.g. GalNAc T1, GalNAc T2, T3, T4, etc.) and UDP-GalNAc. Also, if needed, galactose can be added using galactosyltransferase and UDP-galactose.

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GlycoPEGylation using sialyltransferase. The glycopeptides (1 mg/mL) bearing terminal galactose in Tris buffered saline + 0.02% sodium azide are incubated with CMP-SA-PEG (0.75 mM) and 0.4 U/mL sialyltransferase (ST3Gal3 or ST3Gal4 for N-glycans on EPO and interferon beta; ST3Gal4, or ST3Gal1 for O-glycans on interferon alpha) at 32°C for 24 hours. Other transferases that may work include the 2,6 sialyltransferase from *Photobacterium damsella*. The acceptor peptide concentration is most preferably in the range of 0.1 mg/mL up to the solubility limit of the peptide. The concentration of CMP-SA-PEG should be sufficient for there to be excess over the available sites, but not so high as to cause peptide solubility problems due to the PEG, and may range from 50 μM up to 5 mM, and the temperature may range from 2°C up to 40°C. The time required for complete reaction will depend on the temperature, the relative amounts of enzyme to acceptor substrate, the donor substrate concentration, and the pH.

## 34. GlycoPEGylation of Interferon a produced in CHO cells

Preparation of Asialo-Interferon α. Interferon alpha produced from CHO cells is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 5 mM CaCl<sub>2</sub> and concentrated to 500 μL in a Centricon Plus 20 centrifugal filter. The solution is incubated with 300 mU/mL Neuraminidase II (*Vibrio cholerae*) for 16 hours at 32 °C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel performed. The reaction mixture is then added to prewashed N-(*p*-aminophenyl)oxamic acid-agarose conjugate (800 μL/mL reaction volume) and the washed beads gently rotated for 24 hours at 4 °C. The mixture is centrifuged at 10,000 rpm and the supernatant was collected. The beads are washed 3 times with Tris-EDTA buffer, once with 0.4 mL Tris-EDTA buffer and once with 0.2 mL of the Tris-EDTA buffer and all supernatants were pooled. The supernatant is dialyzed at 4 °C against 50 mM Tris –HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub> and then twice more against 50 mM Tris –HCl pH 7.4, 1 M

NaCl, 0.05% NaN<sub>3</sub>. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter and stored at -20 °C. The conditions for the IEF gel are run according to the procedures and reagents provided by Invitrogen. Samples of native and desialylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

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Preparation of Interferon-alpha-(alpha2,3)-Sialyl-PEG. Desialylated interferonalpha is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of ST3Gal1 at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and desialylated Interferon-alpha are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Interferon-alpha-(alpha2,8)-Sialyl-PEG. Interferon-alpha produced in CHO, which contains an alpha2,3-sialylated O-linked glycan, is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN3, pH 7.2. The solution is incubated 20 with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of CST-II at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction has CMP-SA-PEGfluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line 25 fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated interferon-alpha are dialyzed against water and analyzed by MALDI-TOF MS. 30

Preparation of Interferon-alpha-(alpha2,6)-Sialyl-PEG. Interferon-alpha, containing only O-linked GalNAc, was dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of ST6GalNAcl or II at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated interferon-alpha are dialyzed against water and analyzed by MALDI-TOF MS.

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# 35. GlycoPEGylation of Interferon-β-1a with PEG (10 kDa) and PEG (20 kDa)

This example illustrates a procedure PEGylate Interferon- $\beta$  with either PEG (10 kDa) or PEG (20 kDa).

Briefly, Interferon- $\beta$ -1a (INF- $\beta$ ) was obtained from Biogen (Avonex<sup>TM</sup>). The IFN- $\beta$  was first purified by Superdex-75 chromatography. The IFN- $\beta$  was then desialylated with *Vibrio cholerae* sialidase. The INF- $\beta$  was then PEGylated with SA-PEG (10 kDa) or SA-PEG (20 kDa) and purified with Superdex-200 chromatography.

Superdex-75 chromatography purification. INF-β (150 μg) was applied to a Superdex-75 column (Amersham Biosciences, Arlington Heights, IL) and eluted with PBS with 0.5 M NaCl, 0.02 Tween-20, 20 mM histidine and 10% glycerol. The eluant was monitored for absorbance at 280 nm (Figure 172A and 172B) and fractions were collected. Peaks 4 and 5 were pooled, concentrated in an Amicon Ultra 15 spin filter (Millipore, Billerica, MA), and the buffer was exchanged to TBS with 5 mM CaCl<sub>2</sub>, 0.02% Tween-20, 20 mM histidine and 10% glycerol.

Sialidase Reaction. The INF-β was then desialydated with Vibrio cholera salidase (70 mU/ml, CALBIOCHEM®, EMD Biosciences, Inc., San Diego, CA) on agarose in TBS

with 5 mM CaCl<sub>2</sub>, 0.02% Tween-20, 20 mM histidine and 10% glycerol. The reaction was carried out at 32°C for 18 hours. The INF-β was removed from the agarose with a 0.22 μm Spin-X<sup>TM</sup> filter (Corning Technology, Inc., Norcross, GA). Figure 173A depicts the MALDI analysis of glycans released from native INF-β. The native INF-β has many glycoforms containing terminal sialic acid moieties. Figure 173B depicts the MALDI analysis of glycans released from desialylated INF-β. The desialylated INF-β has primarily one glycoform which is bi-antennary with terminal galactose moieties.

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Lectin Dot-Blot Analysis of Sialylation. Samples of the INF-β from the desialidase reaction were dot-blotted onto nitrocellulose and then blocked with Tris buffered saline (TBS: 0.05M Tris, 0.15M NaCl, pH 7.5) and DIG kit (glycan differentiation kit available from Roche #1 210 238) blocking buffer. Some of the blots were incubated with *Maackia amurensis* agglutinin (MAA) labeled with digoxogenin (DIG) (Roche Applied Science, Indianapolis, IL) to detect α2,3-sialylation of INF-β. These blots were washed with TBS then incubated with anti-digitonin antibody labeled with alkaline phosphatase, then washed again with TBS and developed withNBT/X-phosphate solution, wherein NBT is 4-nitro blue tetrazolium chloride and X-phosphate is 5-bromo-4-chloro—3indoyl phosphate. The left side of Figure 174 depicts the results of the MAA blot of INF-β after the desialylation reaction. The INF-β is partially disialylated, as indicated by the decrease in dot development as compared to native INF-β in the desialylated samples.

Other blots were incubated with *Erthrina cristagalli* lectin (ECL) labeled with biotin (Vector Laboratories, Burlingame, CA) to detect exposed galactose residues on INF- $\beta$ . After incubation with 2.5 µg/ml ECL, the blots were washed in TBS and incubated with streptavidin labeled with alkaline phosphatase. The blots were then washed again and developed. The right side of Figure 174 depicts the ECL blot after development. The increased intensity of the dot of desialylated INF- $\beta$  as compared to the native INF- $\beta$  indicate more exposed galactose moieties and therefore extensive desialylation.

PEGylation of Desialylated INF- $\beta$  with SA-PEG (10 kDa). Desialylated INF- $\beta$  (0.05 mg/ml) was PEGylated with ST3Gal3 (50 mU/ml) and CMP-SA-PEG (10 kDa) (250  $\mu$ M) in an appropriate buffer of TBS + 5 mM CaCl<sub>2</sub>, 0.02% Tween 20, 20 mM histidine, 10%

glycerol for 50 hours at 32°C. Figure 175 depicts the SDS-PAGE analysis of the reaction products showing PEGylated INF- $\beta$  at approximately 98 kDa.

PEGylation of Desialylated INF-β with SA-PEG (20 kDa). Desialylated INF-β (0.5 mg/ml) was PEGylated with ST3Gal3 (170 mU/ml) and CMP-SA-PEG (20 kDa) in an appropriate buffer of TBS + 5 mM CaCl<sub>2</sub>, 0.02% Tween 20, 20 mM histidine, 10% glycerol for 50 hours at 32°C. Figure 176 depicts the SDS-PAGE analysis the products of the PEGylation reaction. The PEGylated INF-β has many higher molecular weight bands not found in the unmodified INF-β indicating extensive PEGylation.

Superdex-200 Purification of INF-β PEGylated with PEG (10 kDa). The products of the PEGylation reaction were separated on a Superdex-200 column (Amersham Biosciences, Arlington Heights, IL) in PBS with 0.5 NaCl, 0.02 Tween-20, 20 mM histidine and 10% glycerol at 1ml/min and 30 cm/hr flow. The eluant was monitored for absorbance at 280 nm (Figure 177) and fractions were collected. Peaks 3 and 4 were pooled and concentrated in an Amicon Ultra 15 spin filter.

# Bioassay of INF- $\beta$ PEGylated with PEG (10 kDa).

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The test is inhibition of the proliferation of the lung carcinoma cell line, A549. The A549 cell line are lung carcinoma adherent cells growing in RPMI + 10% FBS at 37°C 5% CO<sub>2</sub>. They can be obtained from ATCC # CCL-185. Wash the cells with 10 ml of PBS and remove the PBS. Add 5 ml of trypsin, incubate for 5 minutes at room temperature or 2 minutes at 37°C. When the cells are detached resuspend into 25 ml of media and count the cells. Dilute the cells at a concentration of 10000 cells/ml and add 200 ul / well (96 wells plate). Incubate for 4 hours at 37°C 5% CO<sub>2</sub>. Prepare 1 ml of IFN B at a concentration of 0.1 ug/ml. Filter it under the hood with a 0.2 um filter. Add 100 ul per well (8 replicates = 1 lane). Incubate for 3 days (do not let the cells go to confluence). Remove 200 ul of media (only 100ul per well left). Add 25  $\mu$ l of MTT (Sigma) (5 mg/ml filtered 0.22 $\mu$ m). Incubate for 4 hours at 37°C and 5% CO<sub>2</sub>. Aspirate the media gently and add 100  $\mu$ l of a mixture of isopropanol (100 ml and 6N HCl. Aspirate up and down to homogenize the crystal violet. Read OD 570nm (remove the background at 630 or 690 nm).

Figure 178 depicts the results of the bioassay of the peaks containing INF- $\beta$  PEGylated with PEG (10 kDa) as eluted from the Superdex-200 column.

Superdex-200 Purification of INF-β PEGylated with PEG (20 kDa). The products of the PEG (20 kDa) PEGylation reaction were separated on a Superdex-200 column (Amersham Biosciences, Arlington Heights, IL) in PBS with 0.5 NaCl, 0.02 Tween-20, 20 mM histidine and 10% glycerol at 1 ml/min flow. The eluant was monitored for absorbance at 280 nm (Figure 179) and fractions were collected. Peak 3 contained most of the INF-β PEGylated with PEG (20 kDa).

## Endotoxin test of INF-β PEGylated with PEG (20 kDa).

Limulus Lysate Test was performed, BioWhittaker # 50-647U

Table 24. Results of the endotoxin test of INF-β PEGylated with PEG (20 kDa).

	Concentration		
INF-β with PEG (20 kDa)	10 EU/ml	0.06 mg/ml′	0.16 EU/μg
INF-β with PEG (20 kDa)	1 EU/ml	$0.07~\mathrm{mg/ml}$	0.014 EU/μg
Native INF-β	40 EU/ml	0.1  mg/ml	$0.4~{ m EU/\mu g}$

# Remicade™

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## 36. GlycoPEGylation of Remicade<sup>TM</sup> antibody

This example sets forth the procedure to glycoPEGylate a recombinant antibody molecule by introducing PEG molecules to the Fc region glycans. Here Remicade™, a TNF-R:IgG Fc region fusion protein, is the exemplary peptide.

Preparation of Remicade<sup>TM</sup>-Gal-PEG (10 kDa). Remicade<sup>TM</sup> is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl<sub>2</sub>, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM UDP-galactose-PEG (10 kDa) and 0.1 U/mL of galactosyltransferase at 32°C for 2 days to introduce the PEG in the Fc region glycans. To monitor the incorporation of galactose, a small aliquot of the reaction has <sup>14</sup>C-galactose-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The fractions containing product are combined, concentrated, buffer

exchanged and then freeze-dried. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

#### <u>Rituxan<sup>TM</sup></u>

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## 37. Glycoconjugation of geldanamycin to Rituxan™

This example sets forth the glycoconjugation of a small molecule, such as geldanamycin, to the Fc region glycans of an antibody produced in CHO cells, such as Rituxan<sup>TM</sup>. Here, the antibody Rituxan<sup>TM</sup> is used, but one of skill in the art will appreciate that the method can be used with many other antibodies.

Preparation of Rituxan<sup>TM</sup>-Gal-linker-geldanamycin. Rituxan<sup>TM</sup> is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl<sub>2</sub>, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 1 mM UDP-galactose-linker-geldanamycin and 0.1 U/mL of galactosyltransferase at 32°C for 2 days to introduce the geldanamycin in the Fc region glycans. To monitor the incorporation of galactose, a small aliquot of the reaction has <sup>14</sup>C-galactose-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The fractions containing product are combined, concentrated, buffer exchanged and then freeze-dried. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

#### Rnase

38. Remodeling high mannose N-glycans to hybrid and complex N-glycans: Bovine pancreatic RNase

This example sets forth the preparation of bovine pancreas RNase with hybrid or complex N-glycans. The high mannose N-linked glycans of the RNase are enzymatically

digested and elaborated to create hybrid N-linked glycans. Additionally, the high mannose N-linked glycans of the RNase are enzymatically digested and elaborated to create complex N-linked glycans.

High mannose structures of N-linked oligosaccharides in glycopeptides can be modified to hybrid or complex forms using the combination of  $\alpha$ -mannosidases and glycosyltransferases. This example summarizes the results in such efforts using a simple N-Glycan as a model substrate.

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Ribonuclease B (RNaseB) purified from bovine pancreas (Sigma) is a glycopeptide consisting of 124 amino acid residues. It has a single potential *N*-glycosylation site modified with high mannose structures. Due to its simplicity and low molecular weight (13.7 kDa to 15.5 kDa), ribonuclease B is a good candidate to demonstrate the feasibility of the *N*-Glycan remodeling from high mannose structures to hybrid or complex *N*-linked oligosaccharides. The MALDI-TOF spectrum of RNaseB (Figure 180A) and HPLC profile for the oligosaccharides cleaved from RNaseB by N-Glycanase (Figure 180B) indicated that, other than a small portion of the non-modified peptide, the majority of *N*-glycosylation sites of the peptide are modified with high mannose oligosaccharides consisting of 5 to 9 mannose residues.

Conversion of high mannose N-Glycans to hybrid N-Glycans. High mannose N-Glycans were converted to hybrid N-Glycans using the combination of  $\alpha 1,2$ -mannosidase, GlcNAcT-I ( $\beta$ -1,2-N-acetyl glucosaminyl transferase), GalT-I ( $\beta$ 1,4-galactosyltransfease) and  $\alpha$ 2,3-sialyltransferase /or  $\alpha$ 2,6-sialyltransferase as shown in Figure 181.

As an example, high mannose structures in RNaseB were successfully converted to hybrid structures.

Man<sub>5</sub>GlcNAc<sub>2</sub>-R was obtained from Man<sub>5-9</sub>GlcNAc<sub>2</sub>-R catalyzed by a single α1,2-mannosidase cloned from *Trichoderma reesei* (Figure 182). RNase B (1 g, about 67 μmol) was incubated at 30°C for 45 hr with 15 mU of the recombinant *T. reesei* α1,2-mannosidase in MES buffer (50 mM, pH 6.5) in a total volume of 10 mL. Man<sub>6-9</sub>GlcNAc<sub>2</sub>-protein structures have been successfully converted to Man<sub>5</sub>GlcNAc<sub>2</sub>-protein with high efficiency by the recombinant mannosidase.

Alternately, Man<sub>5</sub>GlcNAc<sub>2</sub>-R was obtained from Man<sub>5-9</sub>GlcNAc<sub>2</sub>-R catalyzed by a single α1,2-mannosidase purified from *Aspergillus saitoi* (Figure 183). RNase B (40 μg, about 2.7 nmol) was incubated at 37°C for 42.5 hr with 25 μU of the commercial *A. saitoi* α1,2-mannosidase (Glyko or CalBioChem) in NaOAC buffer (100 mM, pH 5.0) in a total volume of 20 μl. Man<sub>6-9</sub>GlcNAc<sub>2</sub>-protein structures were successfully converted to Man<sub>5</sub>GlcNAc<sub>2</sub>-protein by the commercially available mannosidase. However, a new peak corresponding to the GlcNAc-protein appears in the spectrum, indicating the possible contamination of endoglycosidase H in the preparation. Although several mammalian alphamannosidases were required to achieve this step, the fungal α1,2-mannosidase was very efficient to remove all α1,2-linked mannose residues.

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GlcNAcT-I then added a GlcNAc residue to the Man<sub>5</sub>GlcNAc<sub>2</sub>-R (Figure 184). The reaction mixture after the *T. reesei* α1,2-mannosidase reaction containing RNase B (600 μg, about 40 nmol) was incubated with non-purified recombinant GlcNAcT-I (34 mU) in MES buffer (50 mM, pH 6.5) containing MnCl<sub>2</sub> (20 mM) and UDP-GlcNAc (5 mM) in a total volume of 400 μl. at 37°C for 42 hr. A GlcNAc residue was quantitatively added to Man<sub>5</sub>GlcNAc<sub>2</sub>-protein by the recombinant GlcNAcT-I.

A Gal residue was then added using GalT 1 (Figure 185). The reaction mixture after the GnT-I reaction containing RNase B (120  $\mu$ g, about 8 nmol) was incubated at 37°C for 20 hr with 3.3 mU of the recombinant GalT-1 in Tris-HCl buffer (100 mM, pH 7.3) containing UDP-Gal (7.5 mM) and MnCl<sub>2</sub> (20 mM) in a total volume of 100  $\mu$ l. A Gal residue was added to about 98% of the GlcNAc-Man<sub>5</sub>GlcNAc<sub>2</sub>-protein by the recombinant GalT 1.

The next step was the addition of a sialic acid using an α2,3-sialyltransferase or an α2,6-sialyltransferase (Figure 186). As an example, ST3Gal III, an α2,3-sialyltransferase was used. The reaction mixture after the GalT-1 reaction containing RNase B (13 μg, about 0.87 nmol) was incubated at 37°C for 16 hr with 8.9 mU of recombinant ST3Gal III in Tris-HCl buffer (100 mM, pH 7.3) containing CMP-Sialic acid (5 mM) and MnCl<sub>2</sub> (20 mM) in a total volume of 20 μl. A sialic acid residue was added to about 90% of the Gal-GlcNAc-Man<sub>5</sub>GlcNAc<sub>2</sub>-protein by recombinant ST3Gal III using CMP-SA as the donor. The yield can be further improved by adjusting the reaction conditions.

For convenience, no purification or dialysis step was required after each reaction described above. More interesting, GalT 1 and ST3Gal III can be combined in a one-pot reaction. Similar yields were obtained as compared with the separate reactions. The reaction mixture after the GlcNAcT-I reaction containing RNase B (60 μg, about 4 nmol) was incubated at 37°C for 20 hr with 1.7 mU of recombinant GalT 1, 9.8 mU of recombinant ST3Gal III in Tris-HCl buffer (100 mM, pH 7.3) containing UDP-Gal (7.5 mM), CMP-sialic acid (5 mM) and MnCl<sub>2</sub> (20 mM) in a total volume of 60 μl.

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As shown in Figure 187, SA-PEG (10 kDa) was successfully added to the RNaseB. The reaction mixture after the GalT-1 reaction containing RNase B (6.7 μg, about 0.45 nmol) was dialyzed against H<sub>2</sub>O for 1 hour at room temperature and incubated at 37°C for 15.5 hours with 55 mU of the recombinant ST3Gal III in Tris-HCl buffer (50 mM, pH 7.3) containing CMP-SA-PEG (10 kDa) (0.25 mM) and MnCl<sub>2</sub> (20 mM) in a total volume of 20 μl. PEG-modified sialic acid residues were successfully added to the Gal-GlcNAc-Man<sub>5</sub>GlcNAc<sub>2</sub>-peptide by the recombinant ST3Gal III. The yield can be further improved by adjusting the reaction conditions.

Conversion of high mannose N-Glycans to complex N-Glycans. To achieve this conversion, a GlcNAcβ1,2Man<sub>3</sub>GlcNAc<sub>2</sub>-peptide intermediate is obtained. As shown in Figure 188, there are at least four feasible routes to carry out the reaction from Man<sub>5</sub>GlcNAc<sub>2</sub>-peptide to this intermediate:

Route I: The Man<sub>5</sub>GlcNAc<sub>2</sub>-peptide produced by the fungal  $\alpha$ 1,2 mannosidase is a substrate of GlcNAc transferase I (GlcNAcT-I, enzyme 2) which adds one GlcNAc. The terminal  $\alpha$ 1,3- and  $\alpha$ 1,6-linked mannose residues of GlcNAcMan<sub>5</sub>GlcNAc<sub>2</sub>-peptide is removed by Golgi  $\alpha$ -mannosidase II (ManII, enzyme 5). This route is a part of the natural pathway for the processing of *N*-linked oligosaccharides carried out in higher organisms.

Route II: Two mannose residues are first removed by an α-mannosidase (enzyme 6), then a GlcNAc is added by GlcNAcT-I (enzyme 2). Other than its natural acceptor Man<sub>5</sub>GlcNAc<sub>2</sub>-R, GlcNAcT-I can also recognize Man<sub>3</sub>GlcNAc<sub>2</sub>-R as its substrate and add one GlcNAc to the mannose core structure to form GlcNAcMan<sub>3</sub>GlcNAc<sub>2</sub>-peptide.

Route III: The  $\alpha$ 1,6-linked mannose is removed by an  $\alpha$ 1,6-mannosidase, followed by the addition of GlcNAc by GlcNAcT-I and removal of the terminal  $\alpha$ 1,3-linked mannose

by an  $\alpha$ 1,3-mannosidase. From the experimental data obtained, GlcNAcT-I can recognize this Man<sub>4</sub>GlcNAc<sub>2</sub>-peptide as acceptor and add one GlcNAc residue to form GlcNAcMan<sub>4</sub>GlcNAc<sub>2</sub>-peptide.

Route IV: Similar to Route III,  $\alpha$ 1,3-linked mannose is removed by an  $\alpha$ 1,3-mannosidase, followed by GlcNAcT-I reaction. Then the terminal  $\alpha$ 1,6-linked mannose can be removed by an  $\alpha$ 1,6-mannosidase.

After the function of GlcNAcT-I (responsible for the addition of the GlcNAc β1,2-linked to the α1,3-mannose on the mannose core) and GlcNAcT-II (responsible for the addition of a second GlcNAc β1,2-linked to the α1,6-mannose on the mannose core), the GlcNAc<sub>2</sub>Man<sub>3</sub>GlcNAc<sub>2</sub>-peptide can be processed by GalT 1 and sialyltransferase to form biantennary complex N- Glycans. Other GlcNAc transferases such as GlcNAcT-IV, GlcNAcT-V, and/or GlcNAcT-VI (Figure 188 and Figure 189) can also glycosylate the GlcNAc<sub>2</sub>Man<sub>3</sub>GlcNAc<sub>2</sub>-peptide. Additional glycosylation by the GalT 1 and sialyltransferases will form multi-antennary complex N-glycans. The enzyme GlcNAcT-III catalyzes the insertion of a bisecting GlcNAc, thus preventing the actions of ManII and subsequent action of transferases GlcNAcT-II, GlcNAcT-IV and GlcNAcT-V.

## Tissue-Type Plasminogen Activator (TPA)

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## 39. Fucosylation of TPA to create Sialyl Lewis X

This example sets forth the preparation of Tissue Tissue-type Plasminogen Activator (TPA) with N-linked sialyl Lewis X antigen.

Sialylation. TPA expressed in mammalian cells will often contain a majority of the glycans terminating in sialic acid, but to ensure complete sialylation, it would be beneficial to first perform an *in vitro* sialylation. TPA in a suitable buffer (most preferably between pH 5.5 and 9, for example Tris buffered saline, pH 7.2) is incubated with CMP sialic acid and sialyltransferase for a time sufficient to convert any glycans lacking sialic acid to sialylated species. Typical conditions would be 1 mg/mL TPA, 3 mM CMP sialic acid, 0.02 U/mL ST3Gal3, 32°C for 24 hours. Microbial growth can be halted either by sterile filtration or the inclusion of 0.02% sodium azide. The TPA concentration is most preferably in the range 0.1 mg/mL up to the solubility limit of the peptide. The concentration of CMP-SA should be

sufficient for there to be excess over the available sites, and might range from  $50~\mu\mathrm{M}$  up to  $50~\mathrm{mM}$ , and the temperature from  $2^{\circ}\mathrm{C}$  up to  $40^{\circ}\mathrm{C}$ . The time required for complete reaction will depend on the temperature, the relative amounts of enzyme to acceptor substrate, the donor substrate concentration, and the pH. Other sialyltransferases that may be capable of adding sialic acid in 2,3 linkage include ST3Gal4; microbial transferases could also be used.

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Fucosylation. Typical conditions for fucosylation would be 1 mg/mL TPA, 3 mM GDP-fucose, 0.02 U/mL FTVI, 5 mM MnCl<sub>2</sub>, 32°C for 24H in Tris buffered saline. Microbial growth can be halted either by sterile filtration or the inclusion of 0.02% sodium azide. The TPA concentration is most preferably in the range 0.1 mg/mL up to the solubility limit of the peptide. The concentration of GDP-fucose should be sufficient for there to be excess over the available sites, and might range from 50 μM up to 50 mM, and the temperature from 2°C up to 40°C. The time required for complete reaction will depend on the temperature, the relative amounts of enzyme to acceptor substrate, the donor substrate concentration, and the pH. Other fucosyltransferases that may be capable of making sialyl Lewis x include FTVII, FTV, FTIII, as well as microbial transferases could also be used.

# 40. Trimming of high mannose to tri-mannose core structure: Tissue-type Plasminogen Activator produced in CHO

This example sets forth the preparation of Tissue-type Plasminogen Activator with a trimannose core by trimming back from a high mannose glycan.

Tissue-type plasminogen activator (TPA) is currently produced in Chinese Hamster Ovary (CHO) cells and contains a low amount of high mannose N-linked oligosaccharide. The mannoses can be trimmed down using a variety of the specific mannosidases. The first step is to generate Man5GlcNAc2(Fuc0-1) from Man9GlcNAc2(Fuc0-1). This can be done using mannosidase I. Then either GlcNAcT1 (GlcNAc transferase I) is used to make GlcNAc1Man5GlcNAc2(Fuc0-1) or Mannosidase III is used to make Man3GlcNAc2(Fuc0-1). From Man3GlcNAc2(Fuc0-1), GlcNAc1Man3GlcNAc2(Fuc0-1) can be produced using GlcNAcT1 or from GlcNAc1Man5GlcNAc2(Fuc0-1), GlcNAc1Man3GlcNAc2(Fuc0-1) can be produced using Mannosidase II. GlcNAc1Man3GlcNAc2(Fuc0-1) is then converted into GlcNAc2Man3GlcNAc2(Fuc0-1) using GlcNAcTransferase II (GlcNAcTII). The two

terminal GlcNAc residues are then galactosylated using GalTI and then sialylated with SA-PEG using ST3GalIII.

Conversely, TPA can be produce in yeast or fungal systems. Similar processing would be required for fungal derived material.

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# 41. Generation and PEGylation of GlcNAc-ASN structures: TPA produced in Yeast

This example sets forth the preparation of PEGylated GlcNAc-Asn structures on a peptide such as TPA expressed in yeast.

Yeast expression is expected to result in a TPA which contains a single N-linked mannan-type structure. This recombinant glycoprotein is first treated with endoglycosidase H to generate GlcNAc structures on the asparagine (Asn) residues on the peptide.

The GlcNAc-Asn structures on the peptide/protein backbone are then be modified with galactose or galactose-PEG using UDP-galactose or UDP-galactose-6-PEG, respectively, and a galactosyltransferase such as GalT1. In one case, the galactose-PEG is the terminal residue. In the second case, the galactose is further modified with SA-PEG using a CMP-SA-PEG donor and a sialyltransferase such as ST3GalIII. In another embodiment, the GlcNAc-Asn structures on the peptide/protein backbone may be galactosylated and sialylated as described above, and then further sialylated using CMP-SA-PEG and an α2,8-sialyltransferase such as the enzyme encoded by the *Campylobacter jejuni* cst-II gene.

#### Transferrin

## 42. GlycoPEGylation of Transferrin

This example sets forth the preparation of asialotransferrin and its sialylation with PEG-CMP-sialic acid.

Preparation of Asialo-transferrin. Human-derived holo-Transferrin, (10 mg) was dissolved in 500 μL of 50 mM NaOAc, 5 mM CaCl<sub>2</sub>, pH 5.5. To this solution was added 500 mU Neuraminidase II (*Vibrio cholerae*) and the reaction mixture was shaken gently for 20.5 hours at 37 °C. The reaction mixture was added to the prewashed N-(*p*-aminophenyl)oxamic acid-agarose conjugate (600 μL) and the washed beads gently rotated

for 24 hours at 4 °C. The mixture was centrifuged at 10,000 rpm and the supernatant was collected. The reaction mixture was adjusted to 5 mM EDTA by addition of 100  $\mu$ L of 30 mM EDTA to the washed beads, which were gently rotated for 20 hours at 4 °C. The suspension was centrifuged for 2 minutes at 10,000 rpm and the supernatant was collected. The beads were washed 5 times with 0.35 mL of 50 mM NaOAc, 5 mM CaCl<sub>2</sub>, 5 mM EDTA, pH 5.5 and all supernatants were pooled. The enzyme solution was dialyzed twice at 4 °C into 15 mM Tris-HCl, 1 M NaCl, pH 7.4. 0.3 mL of the transferrin solution (3.3 mL total) was removed and dialyzed twice against water. The remainder was dialyzed twice more at 4 °C against phosphate buffered saline. The dialyzed solution was stored at –20 ° C. Protein samples were analyzed by IEF Electrophoresis. Samples (9  $\mu$ L, 25  $\mu$ g) were diluted with 16  $\mu$ L Tris buffer and mixed with 25  $\mu$ L of the sample loading buffer and applied to Isoelectric Focusing Gels (pH 3-7). Gels were run and fixed using standard procedures. Gels were stained with Colloidal Blue Stain.

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Sialyl-PEGylation of asialo-Transferrin. Desialylated transferrin (250 μg) and CMP-sialic acid or CMP-SA-PEG (1 kDa or 10 kDa)(0.05 μmol) were dissolved in 69 μL 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN3, pH 7.2 in 1.5 mL plastic tubes. The tubes were vortexed briefly and 100 mU ST3Gal3 (90 μ L) were added (total volume 250 μ L). The tubes were vortexed again and mixed gently for 24 hours at 32 °C. The reactions were stopped by freezing at -80 °C. Novex Tris-Glycine 8-16% 1 mm gels were used for SDS PAGE analysis (Figure 190). Samples (25 μL, 25 μg) were mixed with 25 μL of sample loading buffer and 0.4 μL of β-mercaptoethanol and heated for 6 minutes at 85 °C. Gels were run using standard conditions and stained with Colloidal Blue Stain. IEF gels were also performed as described above Figure 191). Samples were also dialyzed against water analyzed by MALDI-TOF.

Results. MALDI was also performed. Native transferrin (78729); asialotransferrin (78197); resialylated transferrin (79626/80703); with SA-PEG 1k (79037 (1); 80961 (2); 82535 (3); 84778 (4)); with SA-PEG 5k (90003 (2); 96117 (3); 96117 (4)); with SA-PEG 10k (100336 (2); 111421 (3); 122510 (4)).

#### 43. Transferrin-GDNF

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This example sets forth the procedures for the glycoconjugation of proteins, and in particular, transferrin is glycoconjugated to GDNF. Transferrin-SA-Linker-Gal-UDP is prepared from transferrin. The galactose residue is removed from GNDF glycans, and Transferrin-SA-Linker-Gal-UDP is conjugated to GNDF glycans using a galactosyltransferase.

Preparation of agalacto-GDNF. GDNF produced in NSO cells (NSO murine myeloma cells) is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, and is incubated with 300 mU/mL beta-galactosidase-agarose conjugate for 16 hours at 32°C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel performed according to Invitrogen procedures. The mixture is centrifuged at 10,000 rpm and the supernatant is collected. The supernatant is dialyzed at 4 °C against 50 mM Tris –HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub> and then twice more against 50 mM Tris –HCl pH 7.4, 1 M NaCl, 0.05% NaN<sub>3</sub>. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter and stored at –20 °C. The conditions for the IEF gel are run according to the procedures and reagents provided by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Transferrin-SA-Linker-Gal-UDP. Asialo-transferrin is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with CMP-sialic acid-linker-Gal-UDP (molar amount to add 1 molar equivalent of nucleotide sugar to transferrin) and 0.1 U/mL of ST3Gal3 at 32°C for 2 days. To monitor the incorporation of sialic acid, a small aliquot of the reaction has <sup>14</sup>C-SA-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

The solution is incubated with 5 mM CMP-sialic acid and 0.1 U/mL of ST3Gal3 (to cap any unreacted transferrin glycans) at 32°C for 2 days. The incorporation into the peptide is quantitated using an in-line UV detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE

and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Transferrin-SA-Linker-Gal-GDNF. The transferrin-SA-Linker-Gal-UDP prepared as described above is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl<sub>2</sub>, 0.05% NaN<sub>3</sub>, pH 7.2. The solution is incubated with 2.5 mg/mL agalacto-GDNF and 0.1 U/mL of galactosyltransferase at 32°C for 2 days. To monitor the incorporation of galactose, a small aliquot of the reaction has <sup>14</sup>C-galactose-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

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When the reaction is complete, the solution is incubated with 5 mM UDP-Gal and 0.1 U/mL of galactosyltransferase (to cap any unreacted transferrin glycans) at 32°C for 2 days followed by addition of 5 mM CMP-SA and 0.1 U/mL of ST3Gal3. After 2 additional days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

The disclosures of each and every patent, patent application, and publication cited herein are hereby incorporated herein by reference in their entirety.

While this invention has been disclosed with reference to specific embodiments, it is apparent that other embodiments and variations of this invention may be devised by others skilled in the art without departing from the true spirit and scope of the invention. The appended claims are intended to be construed to include all such embodiments and equivalent variations.

What is claimed:

1. A cell-free, in vitro method of remodeling a peptide comprising poly(ethylene glycol), the peptide having the formula:

$$\xi$$
—AA— $X^1$ — $X^2$ 

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AA is a terminal or internal amino acid residue of the peptide;

X1-X2 is a saccharide covalently linked to the AA, wherein

X1 is a first glycosyl residue; and

 $X^2$  is a second glycosyl residue covalently linked to  $X^1$ , wherein  $X^1$  and  $X^2$  are selected from monosaccharyl and oligosaccharyl residues;

the method comprising:

- (a) removing  $X^2$  or a saccharyl subunit thereof from the peptide, thereby forming a truncated glycan.
- 15 2. The method according to claim 1 wherein said truncated glycan is formed by removing a Sia residue.
  - 3. The method according to claim 1 wherein said peptide has the formula:

$$\begin{array}{c} (X^{17})_{x} \\ \text{Man} & (X^{3})_{a} \\ \\ \xrightarrow{5} - \text{AA} - \text{GIcNAc} - \text{GIcNAc} - \text{Man} - (X^{4})_{b} \\ \\ \text{Man} - (X^{5})_{c} \\ \\ (X^{7})_{e} \end{array}$$

20 wherein

$X^3, X^4, X^5, X^6, X^7$ , and $X^{17}$ , are independently selected monosaccharyl or		
oligosaccharyl residues; and		
a, b, c, d, e, and x are independently selected from the integers 0, 1 and 2.		

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The method according to claim 3 wherein said oligosaccharyl residue 4. is a member selected from GlcNAc-Gal-Sia and GlcNAc-Gal.

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The method according to claim 3 wherein at least one member selected from a, b, c, d, e and x is 1 or 2.

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The method of claim 3, wherein said removing of step (a) produces a 6. truncated glycan in which at least one of a, b, c, e and x are 0.

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The method of claim 6, wherein X3, X5 and X7 are members 7. independently selected from (mannose)z and (mannose)z-(X8)

wherein

X8 is a glycosyl moiety selected from mono- and oligo-saccharides; and z is an integer between 1 and 20, wherein when z is 3 or greater, each (mannose)z is independently selected from linear

20 and branched structures.

> The method of claim 6 wherein X4 is selected from the group 8. consisting of GlcNAc and xylose.

25

The method of claim 6, wherein X3, X5 and X7 are (mannose)u 9. wherein

u is selected from the integers between 1 and 20, and when u is 3 or greater, each (mannose), is independently selected from linear and branched structures.

30

The method according to claim 3 wherein said peptide has the formula: 10.

wherein

r, s, and t are integers independently selected from 0 and 1.

5

11. The method of claim 1, wherein said peptide has the formula:

wherein

 $X^9$  and  $X^{10}$  are independently selected monosaccharyl or oligosaccharyl

10 residues; and

m, n and f are integers independently selected from 0 and 1.

12. The method of claim 11, wherein said peptide has the formula:

15

wherein

X<sup>16</sup> is a member selected from:

s and i are integers independently selected from 0 and 1.

13. The method of claim 12, wherein said peptide has the formula:

wherein

 $X^{13}$ ,  $X^{14}$ , and  $X^{15}$  are independently selected glycosyl residues; and g, h, i, j, k, and p are independently selected from the integers 0 and 1

10

5

14. The method according to claim 13 wherein at least one of g, h, i, j, k and p is 1.

15

- 15. The method of claim 13, wherein  $X^{14}$  and  $X^{15}$  are members independently selected from GlcNAc and Sia; and i and k are independently selected from the integers 0 and 1.
- 16. The method according to claim 15 wherein at least one of i and k is 1, and if k is 1, g, h, and j are 0.

20

- 17. The method according to claim 1, further comprising:
- (b) contacting the truncated glycan with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer the at least one glycosyl

20

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donor to the truncated glycan, thereby remodeling said peptide comprising poly(ethylene glycol).

- The method according to claim 17 wherein said glycosyl donor
   comprises a modifying group covalently linked thereto.
  - 19. The method of claim 1, further comprising:
  - (c) removing X<sup>1</sup>, thereby exposing AA.
- 10 20. The method according to claim 19, further comprising:
  - (d) contacting AA with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer said at least one glycosyl donor to AA, thereby remodeling said peptide comprising poly(ethylene glycol).
- 15 21. The method according to claim 20 wherein said at least one glycosyl donor comprises a modifying group covalently linked thereto.
  - 22. The method according to claim 21 wherein said modifying group is poly(ethylene glycol).
  - 23. The method according to claim 22 wherein said poly(ethylene glycol) has a molecular weight distribution that is essentially homodisperse.
    - 24. The method of claim 17, further comprising:
- 25 (e) prior to step (b), removing a group added to said saccharide during post-translational modification.
  - 25. The method of claim 24 wherein said group is a member selected from phosphate, sulfate, carboxylate and esters thereof.
    - 26. The method of claim 1 wherein said peptide has the formula:

$$\xi$$
—AA—Z— $X^1$ — $X^2$ 

wherein

Z is a member selected from O, S, NH and a cross-linker.

27. The method of claim 1, wherein said peptide has the formula:

wherein

 $X^{11}$  and  $X^{12}$  are independently selected glycosyl moieties; and r and x are integers independently selected from 0 and 1.

10

5

- 28. The method of claim 27, wherein  $X^{11}$  and  $X^{12}$  are (mannose)<sub>q</sub>, wherein q is selected from the integers between 1 and 20, and when q is three or greater, (mannose)<sub>q</sub> is selected from linear and branched structures.
- 29. A pharmaceutical composition comprising a pharmaceutically acceptable diluent and a remodeled peptide according to claim 1.
  - 30. A cell-free, in vitro method of remodeling a peptide comprising poly(ethylene glycol), said peptide having the formula:

$$\xi$$
 AA- $\left(X^{1}\right)_{u}$ 

20

wherein

AA is a terminal or internal amino acid residue of said peptide;

X¹ is a glycosyl residue covalently linked to said AA, selected from monosaccharyl and oligosaccharyl residues; and

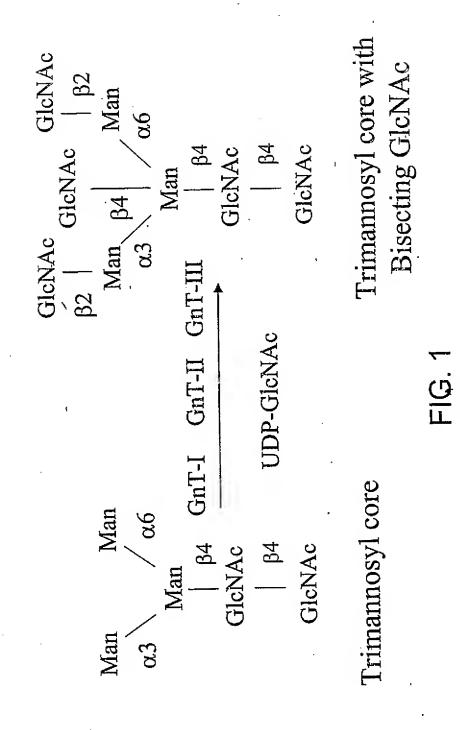
u is an integer selected from 0 and 1,

5 said method comprising:

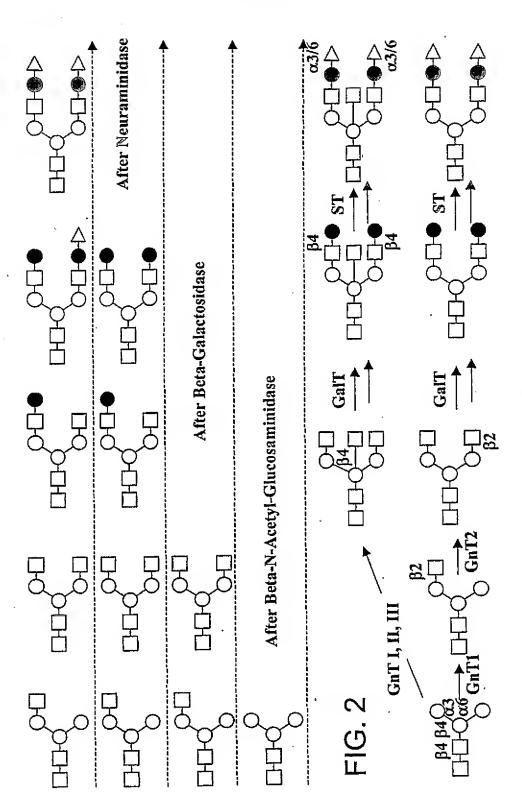
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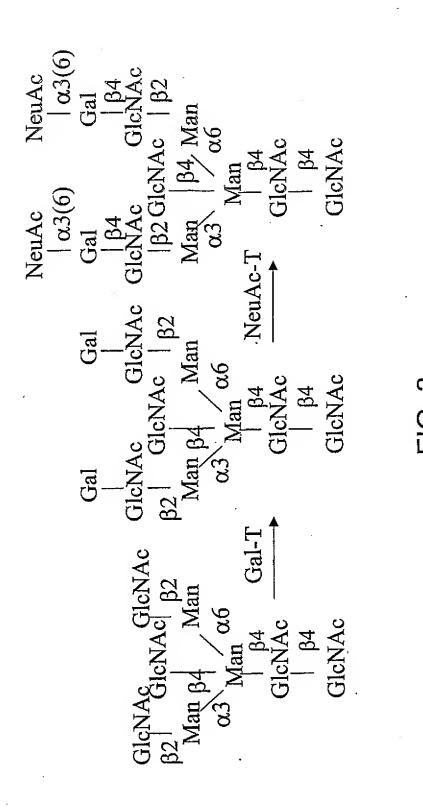
contacting said peptide with at least one glycosyltransferase and at least one glycosyl donor under conditions suitable to transfer said at least one glycosyl donor to said truncated glycan, thereby remodeling said peptide.

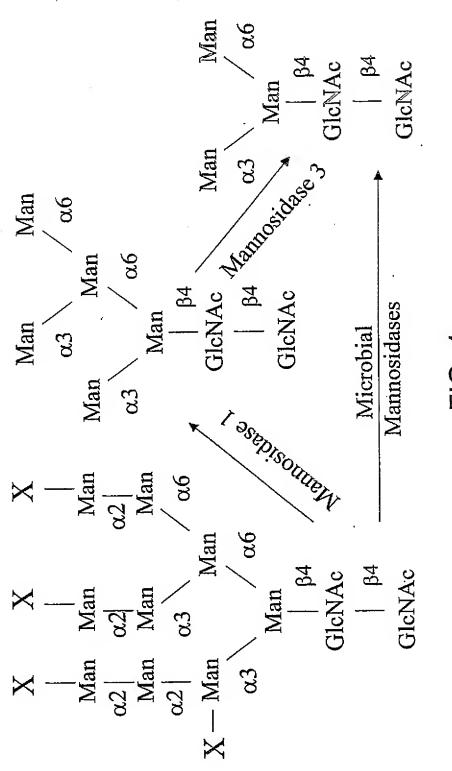
- 10 31. The method according to claim 30 wherein said at least one glycosyl donor comprises a modifying group covalently linked thereto.
  - 32. The method according to claim 30 wherein said modifying group is poly(ethylene glycol).
  - 33. The method according to claim 32 wherein said poly(ethylene glycol) has a molecular weight distribution that is essentially homodisperse.
- 34. A pharmaceutical composition comprising a pharmaceutically
   acceptable diluent and a remodeled peptide according to claim 30.



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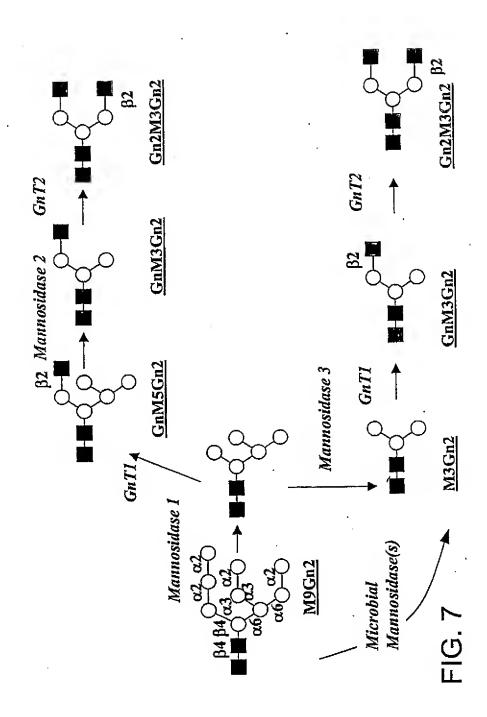


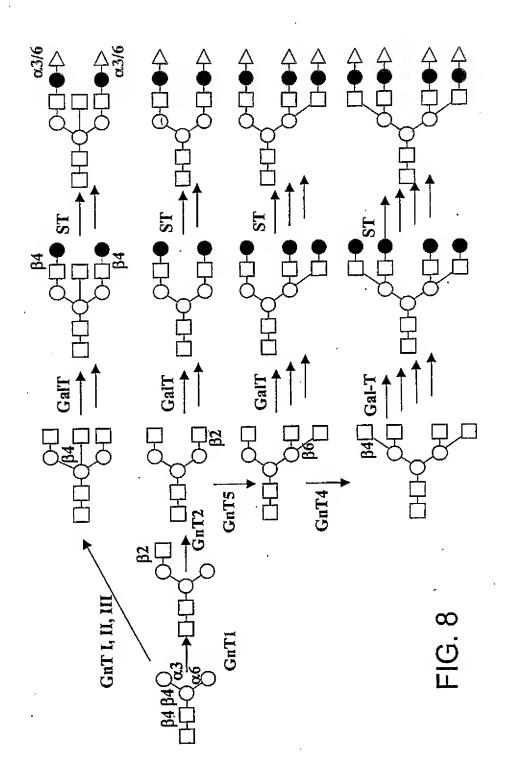




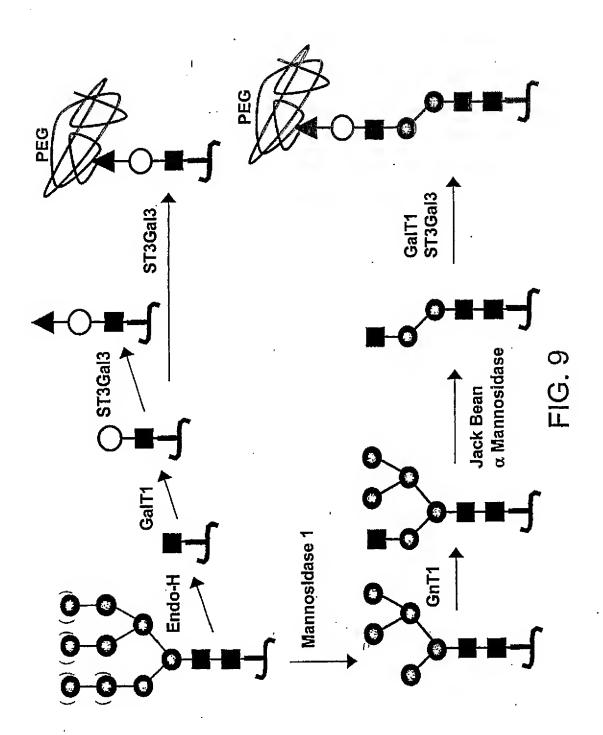
- G. 4

$$(GlcNAc) \qquad (GlcNAc) \qquad (GlcNAc) \\ \beta 2 \qquad \qquad \beta 2 \qquad \beta 3 \qquad \beta 4 \qquad$$

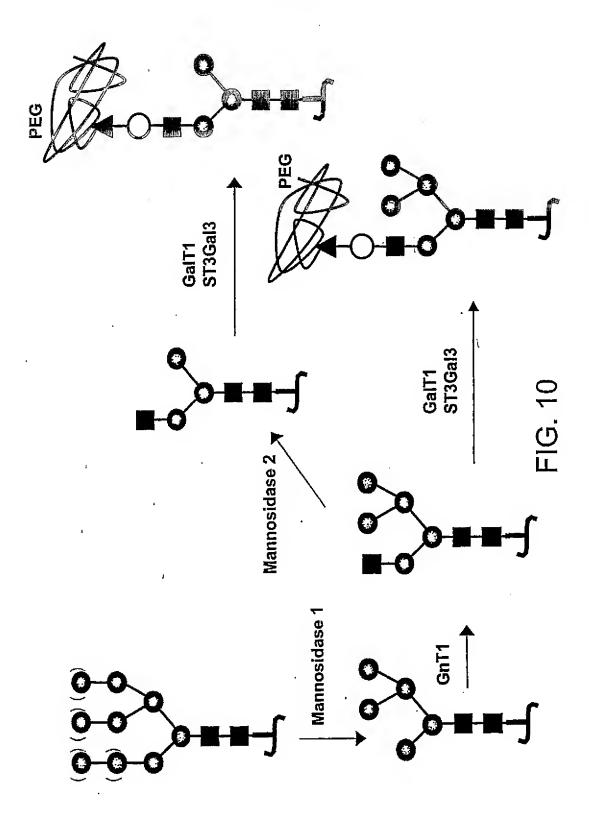


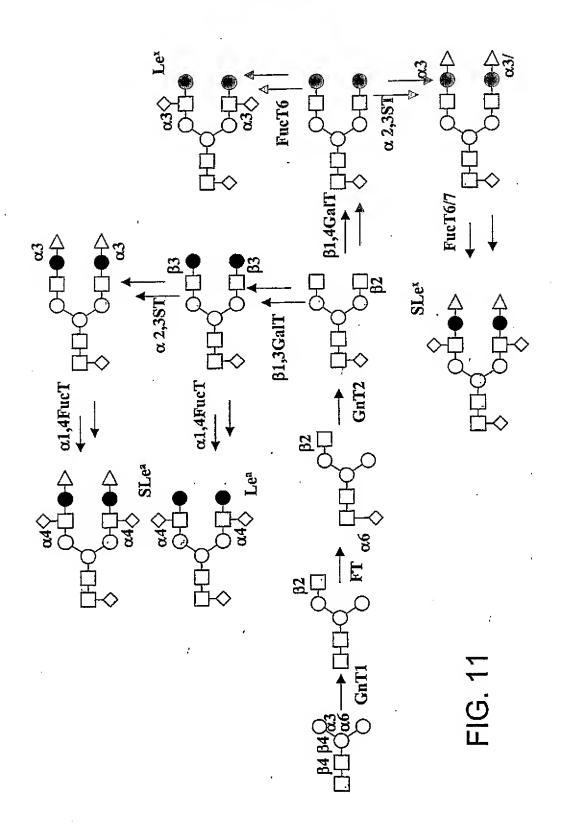


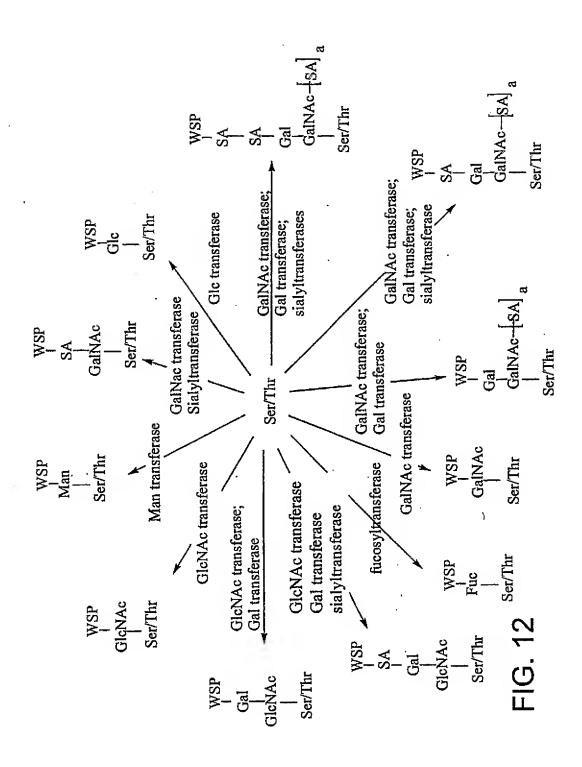
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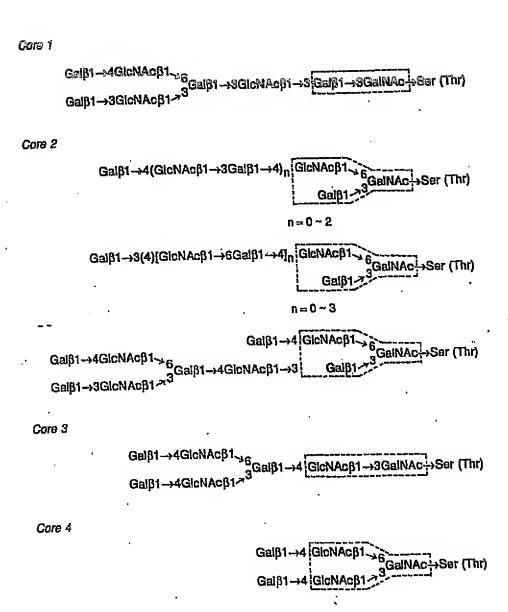
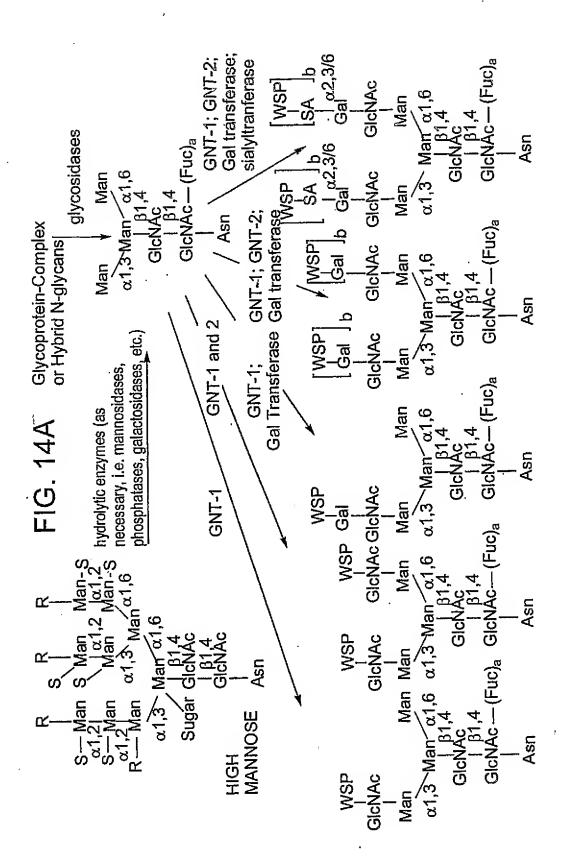
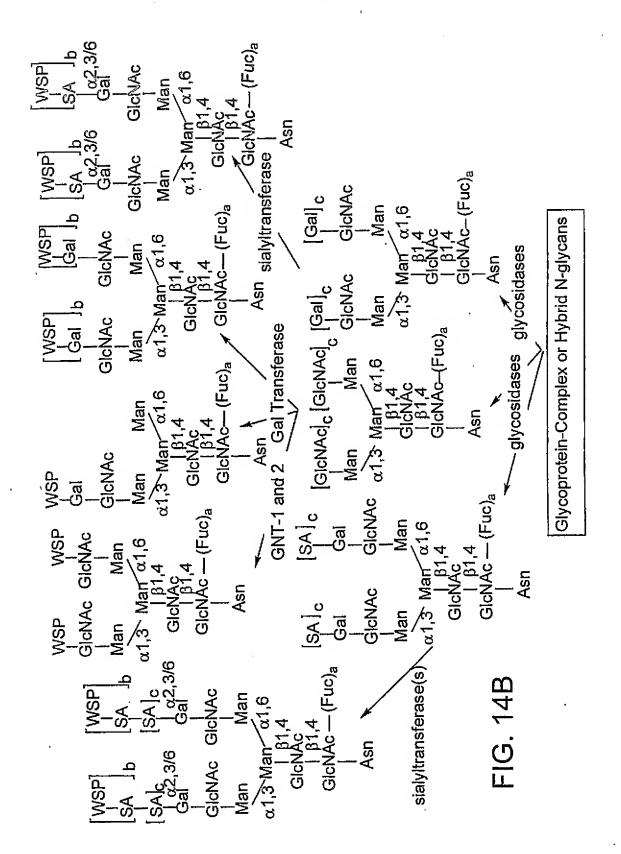


FIG. 13





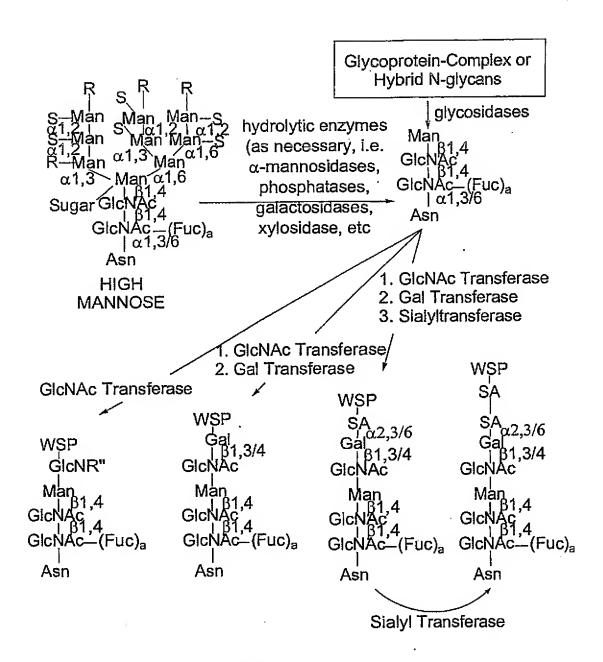


FIG. 15

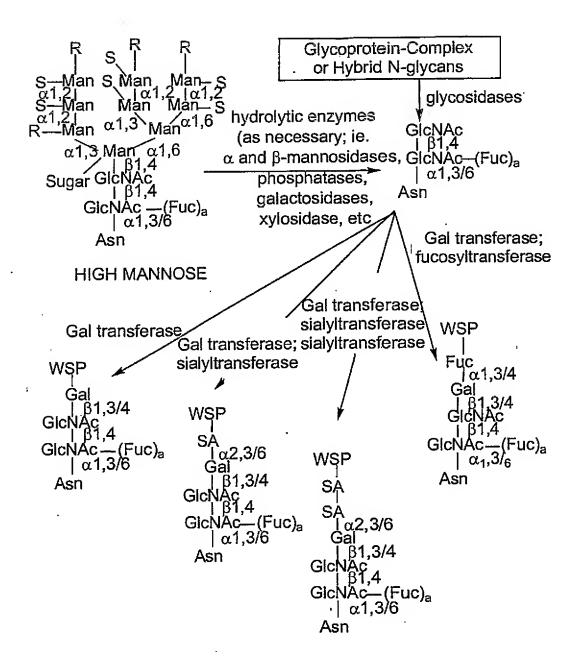


FIG. 16

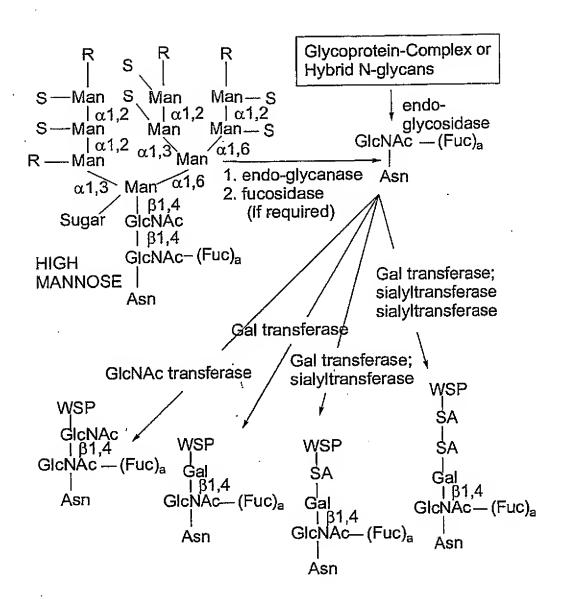
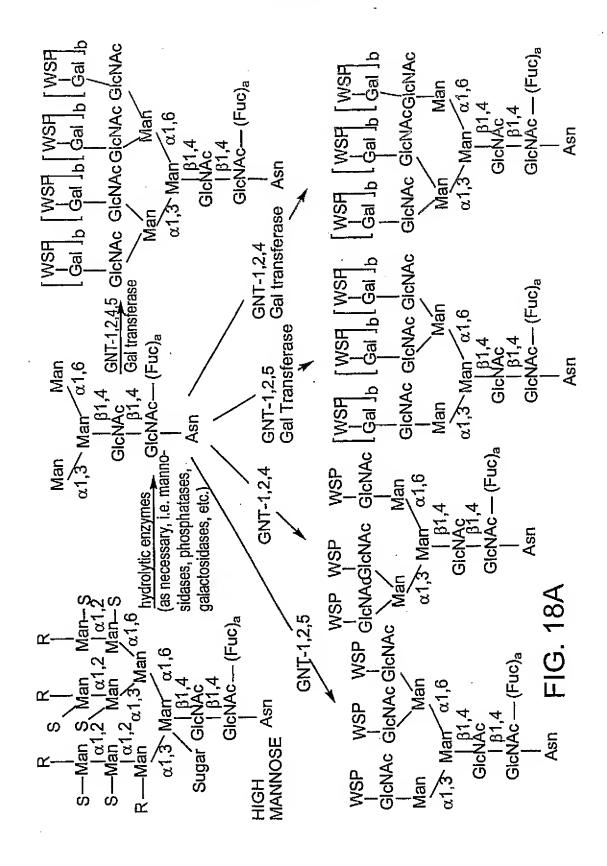
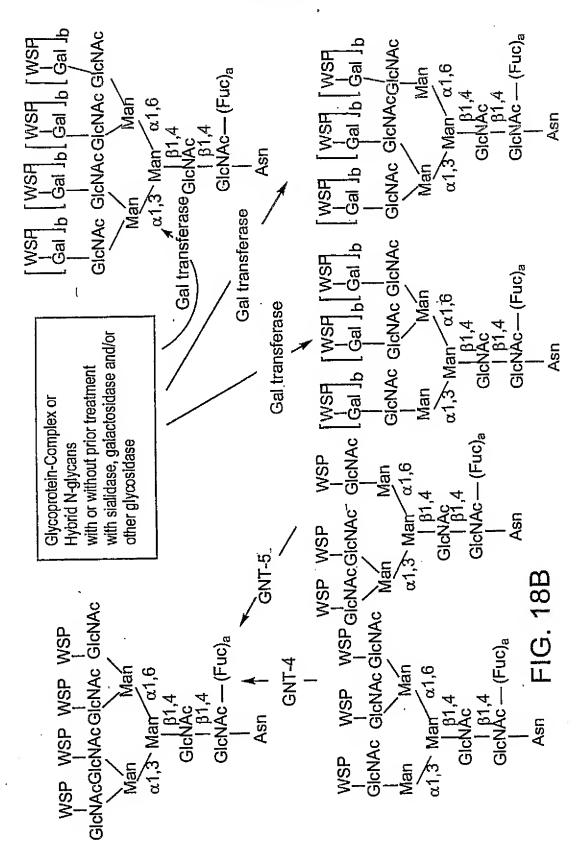
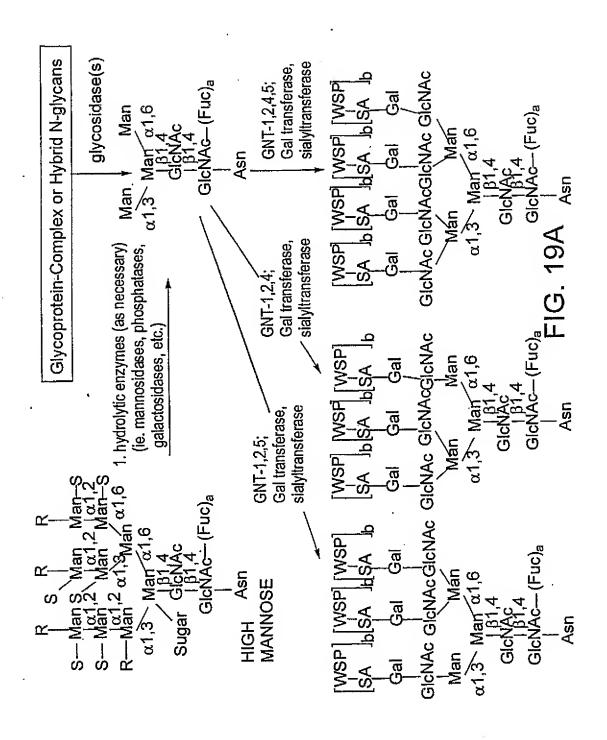
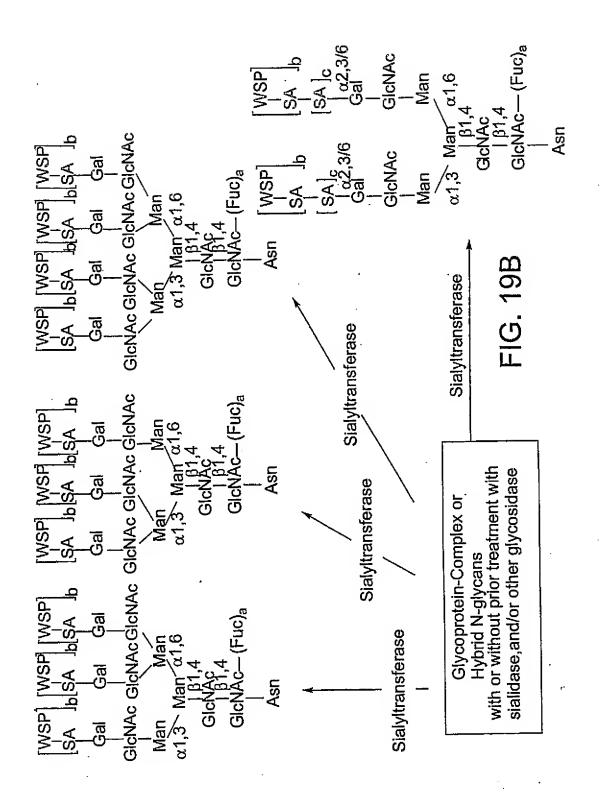


FIG. 17









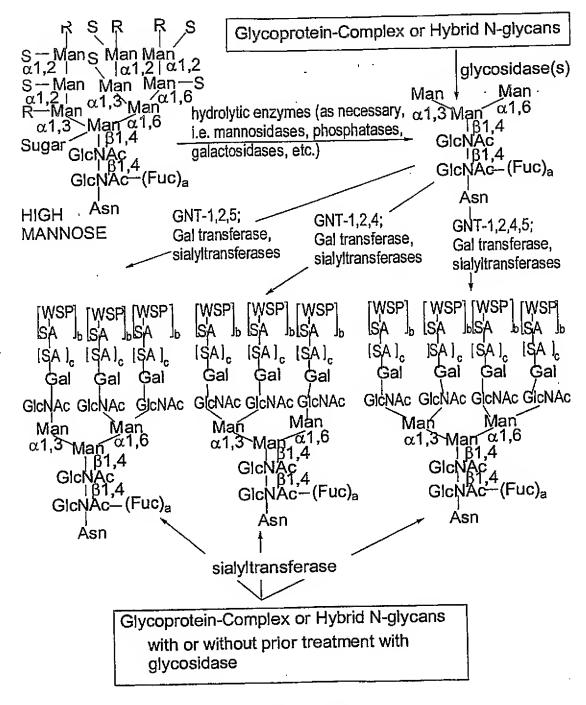
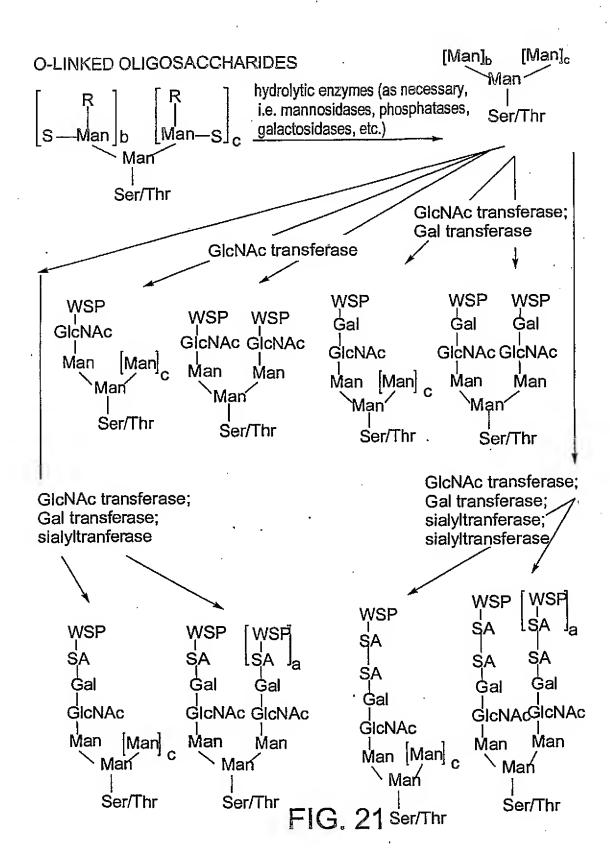
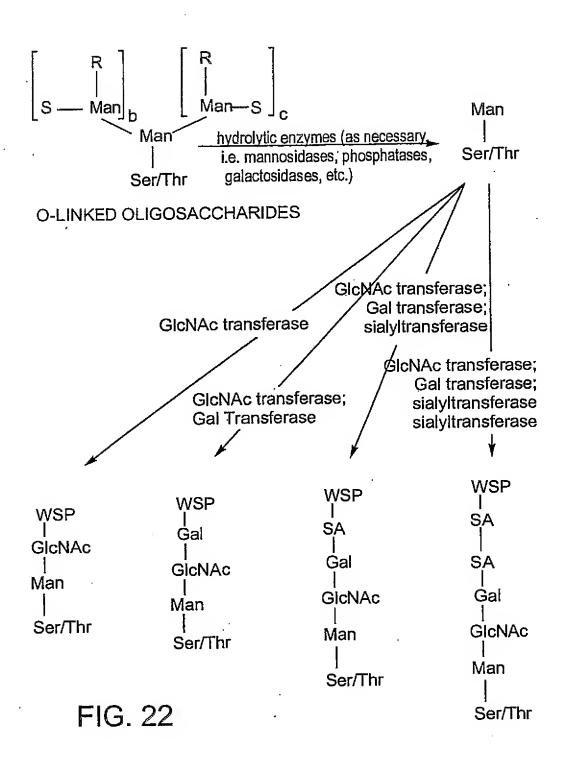
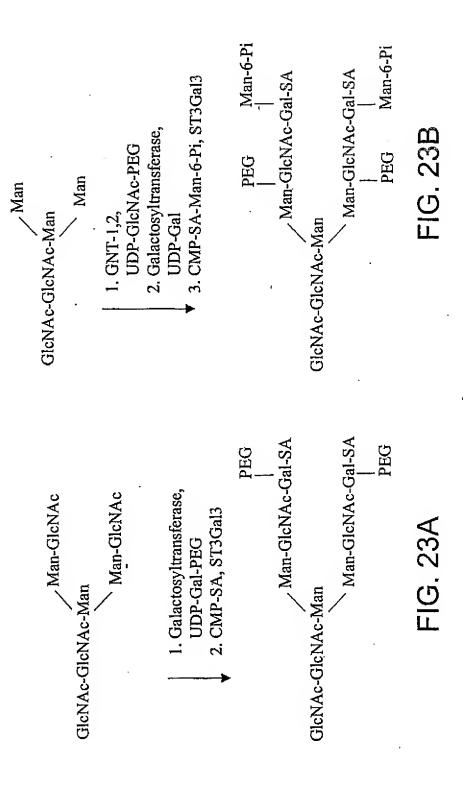
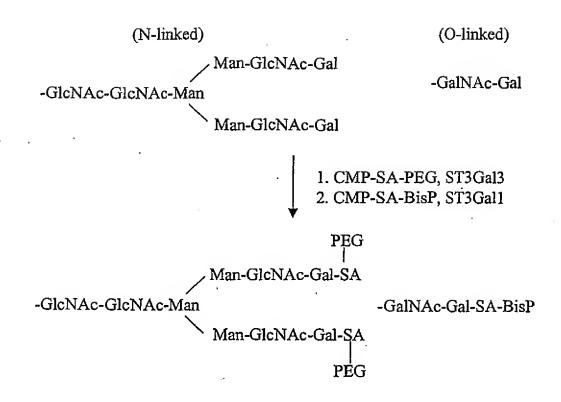


FIG. 20









BisP =Linker-HN-CH(PO<sub>3</sub>),

FIG. 23C

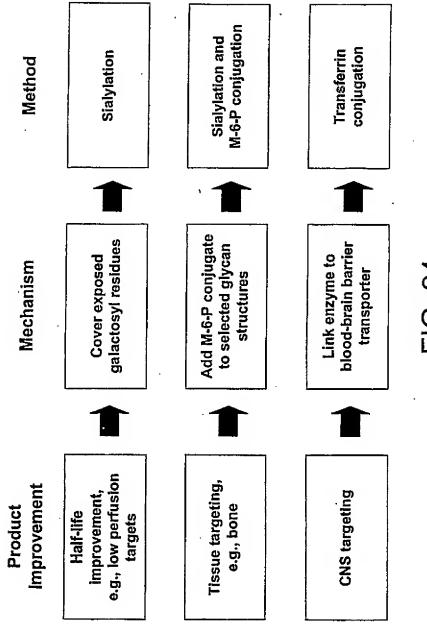
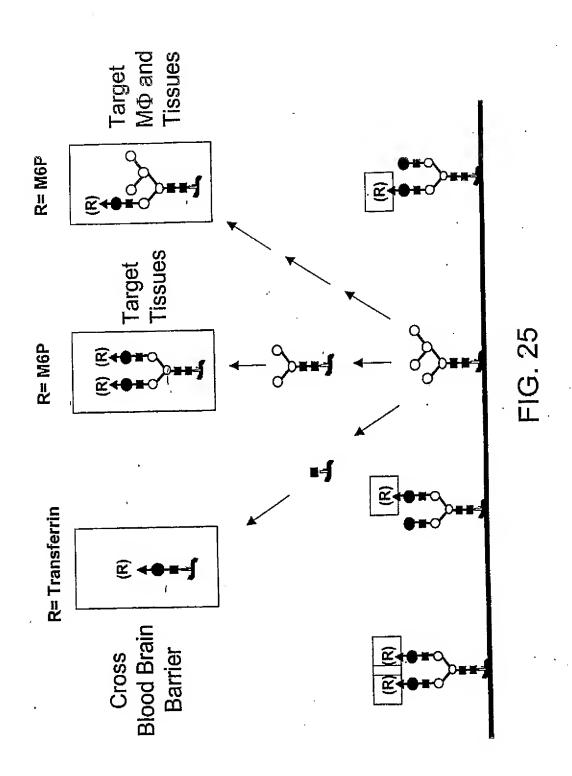
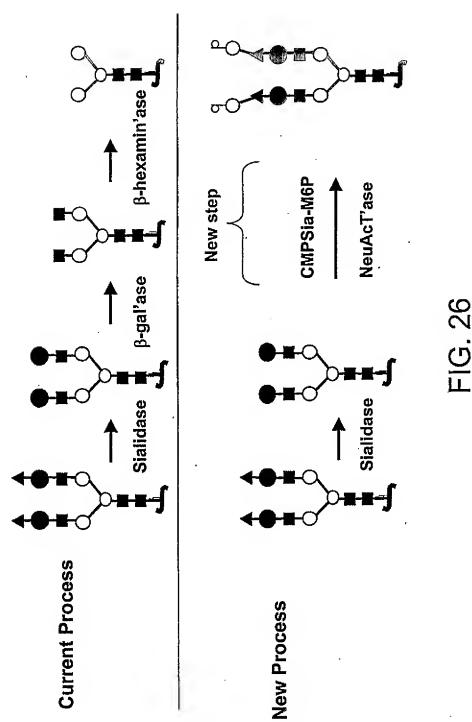
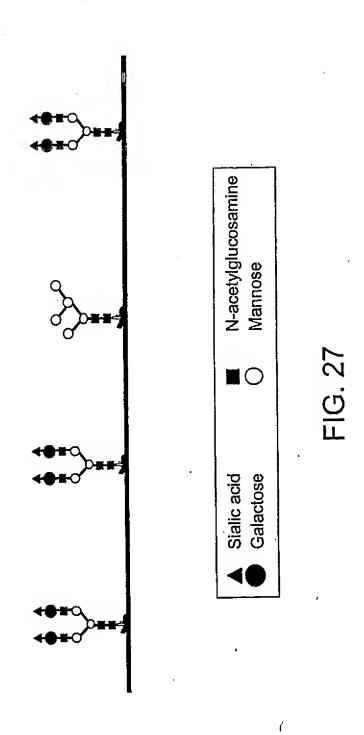


FIG. 24







Al-201 - Autolmmune 12AP1/E5 -- Viventia Biotech Al-301 - Autolmmune 1964 -- Aventis AIDS vaccine - ANRS, CIBG, Hesed 20K growth hormone -- AMUR Biomed, Hollis-Eden, Rome, United 28P6/E6 -- Viventia Biotech Biomedical, American Home Products, 3-Hydroxyphthaloyl-beta-lactoglobulin -Maxygen 4-IBB ligand gene therapy -64-Cu MAb conjugate TETA-1A3 -airway receptor ligand - IC Innovations Mallinckrodt Institute of Radiology AJvW 2 – Ajinomoto AK 30 NGF -- Alkermes 64-Cu MAb conjugate TETA-cT84.66 Albuferon -- Human Genome Sciences 64-Cu Trastuzumab TETA conjugate albumin - Biogen, DSM Anti-Infectives, Genentech Genzyme Transgenics, PPL Therapeutics, A 200 -- Amgen TranXenoGen, Welfide Corp. A10255 - Eli Liliy aldesleukin -- Chiron A1PDX - Hedral Therapeutics alefacept -- Biogen A6 -- Angstrom Alemtuzumab aaAT-III -- Genzyme Allergy therapy -- ALK-Abello/Maxygen, Abciximab - Centocor ALK-Abello/RP Scherer ABI.001 - Atlantic BioPharmaceuticals allergy vaccines -- Allergy Therapeutics ABT-828 - Abbott Alnidofibatide -- Aventis Pasteur Accutin Alnorine -- SRC VB VECTOR Actinohivin ALP 242 -- Gruenenthal activin - Biotech Australia, Human Alpha antitrypsin -- Arriva/Hyland Therapeutics, Curis Immuno/ProMetic/Protease Sciences AD 439 - Tanox Alpha-1 antitrypsin - Cutter, Bayer, PPL AD 519 - Tanox Therapeutics, Profile, ZymoGenetics, Adalimumab -- Cambridge Antibody Tech. Arriva Adenocarcinoma vaccine - Biomira - NIS Alpha-1 protease inhibitor -- Genzyme Adenosine deanimase -- Enzond Transgenics, Welfide Corp. Adenosine A2B receptor antagonists --Alpha-galactose fusion protein -Adenosine Therapeutics **Immunomedics** ADP-001 – Axis Genetics Alpha-galactosidase A -- Research AF 13948 – Affymax Corporation Technologies, Genzyme Afelimomab – Knoll Alpha-glucosidase - Genzyme, Novazyme AFP-SCAN - Immunomedics Alpha-lactalbumin AG 2195 - Corixa agalsidase alfa -- Transkaryotic Therapies Alpha-L-iduronidase -- Transkaryotic Therapies, BioMarin agalsidase beta -- Genzyme alteplase -- Genentech AGENT- Antisoma alvircept sudotox -- NIH Al 300 - Autolmmune ALX1-11 -sNPS Pharmaceuticals Al-101 - Teva Alzheimer's disease gene therapy Al-102 - Teva

#### **FIG. 28A**

AM-133 AMRAD	Anti-angiogenesis monoclonal antibodies -
Amb a 1 immunostim conj. – Dynavax	KS Biomedix/Schering AG
AMD 3100 - AnorMED NIS	Anti-B4 MAb-DC1 conjugate ImmunoGen
AMD 3465 – AnorMED NIS	Anti-B7 antibody PRIMATIZED - IDEC
AMD 3465 – AnorMED NIS	Anti-B7-1 MAb 16-10A1
AMD Fab Genentech	Anti-B7-1 MAb 1G10
Amediplase – Menarini, Novartis	Anti-B7-2 MAb GL-1
AM-F9	Anti-B7-2-gelonin immunotoxin –
Amoebiasis vaccine	Antibacterials/antifungals
Amphiregulin Octagene	Diversa/IntraBiotics
anakinra – Amgen	Anti-beta-amyloid monoclonal antibodies
analgesic Nobex	Cambridge Antibody Tech., Wyeth-Ayerst
ancestim Amgen	Anti-BLyS antibodies Cambridge
AnergiX.RA – Corixa, Organon	Antibody Tech. /Human Genome Sciences
Angiocidin – InKine	Antibody-drug conjugates Seattle
angiogenesis inhibitors ILEX	Genetics/Eos
AngioMab – Antisoma	Anti-C5 MAb BB5-1 Alexion
Angiopoietins Regeneron/Procter &	Anti-C5 MAb N19-8 Alexion
Gamble	Anti-C8 MAb
angiostatin EntreMed	anticancer cytokines BioPulse
Angiostatin/endostatin gene therapy	anticancer matrix – Telios Integra
Genetix Pharmaceuticals	Anticancer monoclonal antibodies – ARIUS,
angiotensin-II, topical Maret	lmmunex
Anthrax - EluSys Therapeutics/US Army	anticancer peptides Maxygen, Micrologix
Medical Research Institute	Anticancer prodrug Tech Alexion
Anthrax vaccine	Antibody Technologies
Anti platelet-derived growth factor D human	anticancer Troy-Bodies Affite Affitech
monocional antibodies CuraGen	anticancer vaccine NIH
Anti-17-1A MAb 3622W94	anticancers - Epimmune
GlaxoSmithKline ,	Anti-CCR5/CXCR4 sheep MAb KS
Anti-2C4 MAb Genentech	Biomedix Holdings
anti-4-1BB monoclonal antibodies Bristol-	
Myers Squibb	Anti-CD11a MAb M17
Anti-Adhesion Platform Tech Cytovax	Anti-CD11a MAb TA-3
Anti-adipocyte MAb Cambridge Antibody	
Tech./ObeSys	Anti-CD11b MAb Pharmacia
antiallergics Maxygen	Anti-CD11b MAb LM2
antiallergy vaccine Acambis	Anti-CD154 MAb Biogen
Anti-alpha-4-integrin MAb	Anti-CD16-anti-CD30 MAb Biotest
Anti-alphavβ3 integrin MAb – Applied	Anti-CD18 MAb — Pharmacia
Molecular Evolution	Anti-CD19 MAb B43 –

FIG. 28B

Anti-CD19 MAb -liposomal sodium butyrate conjugate –	Anti-CD4 MAb 4162W94 — GlaxoSmithKline Anti-CD4 MAb B-F5 — Diaclone
Anti-CD147	Anti-CD4 MAb GK1-5
Anti-CD19 MAb-saporin conjugate –	Anti-CD4 MAb KT6
Anti-CD19-dsFv-PE38-immunotoxin –	
Anti-CD2 MAb 12-15 –	Anti-CD4 MAb CX38
	Anti-CD4 MAb PAP conjugate Bristol-
Anti-CD2 MAb B-E2 Diadone	Myers Squibb
Anti-CD2 MAb OX34 –	Anti-CD4 MAb RIB 5-2
Anti-CD2 MAb OX54 –	Anti-CD4 MAb W3/25
Anti-CD2 MAb OX55 –	Anti-CD4 MAb YTA 3.1.2
Anti-CD2 MAb RM2-1	Anti-CD4 MAb YTS 177-9
Anti-CD2 MAb RM2-2	Anti-CD40 ligand MAb 5c8 Biogen
Anti-CD2 MAb RM2-4	Anti-CD40 MAb
Anti-CD20 MAb BCA B20	Anti-CD40 MAb 5D12 – Tanox
Anti-CD20-anti-Fc alpha RI bispecific MAb -	
Medarex, Tenovus	Anti-CD44 MAb GKWA3
Anti-CD22 MAb-saporin-6 complex –	Anti-CD44 MAb IM7
Anti-CD3 immunotoxin –	Anti-CD44 MAb KM81
Anti-CD3 MAb 145-2C11 Pharming	Anti-CD44 variant monoclonal antibodies
Anti-CD3 MAb CD4lgG conjugate	Corixa/Hebrew University
Genentech	Anti-CD45 MAb BC8-I-131
Anti-CD3 MAb humanised – Protein Design,	
RW Johnson	Anti-CD48 MAb HuLy-m3
Anti-CD3 MAb WT32	Anti-CD48 MAb WM-63
Anti-CD3 MAb-ricin-chain-A conjugate -	Anti-CD5 MAb Becton Dickinson
Anti-CD3 MAb-xanthine-oxidase conjugate	Anti-CD5 MAb OX19
<del>-</del>	Anti-CD6 MAb
Anti-CD30 MAb BerH2 Medac	Anti-CD7 MAb-PAP conjugate
Anti-CD30 MAb-saporin conjugate	Anti-CD7 MAb-ricin-chain-A conjugate
Anti-CD30-scFv-ETA'-immunotoxin	Anti-CD8 MAb – Amerimmune, Cytodyn,
Anti-CD38 MAb AT13/5	Becton Dickinson
Anti-CD38 MAb-saporin conjugate	Anti-CD8 MAb 2-43
Anti-CD3-anti-CD19 bispecific MAb	Anti-CD8 MAb OX8
Anti-CD3-anti-EGFR MAb	Anti-CD80 MAb P16C10 IDEC
Anti-CD3-anti-interleukin-2-receptor MAb	Anti-CD80 MAb P7C10 ID Vaccine
Anti-CD3-anti-MOv18 MAb — Centocor	Anti-CD8-idarubicin conjugate
Anti-CD3-anti-SCLC bispecific MAb	Anti-CEA MAb CE-25
Anti-CD4 idiotype vaccine	Anti-CEA MAb MN 14 – Immunomedics
	Anti-CEA MAb MN14-PE40 conjugate –
Pharmaceuticals, Xenova Group	Immunomedics
Anti-CD4 MAb 16H5	

FIG. 28C

Anti-CEA MAb T84.66-interleukin-2 Anti-heparanase human monoclonal antibodies - Oxford conjugate Glycosciences/Medarex Anti-CEA sheep MAb -- KS Biomedix Anti-hepatitis C virus human monoclonal Holdings antibodies -- XTL Biopharmaceuticals Anti-cell surface monoclonal antibodies -Anti-HER-2 antibody gene therapy Cambridge Antibody Tech. /Pharmacia Anti-c-erbB2-anti-CD3 bifunctional MAb --Anti-herpes antibody - Epicyte Anti-HIV antibody -- Epicyte Otsuka anti-HIV catalytic antibody -- Hesed Biomed Anti-CMV MAb -- Scotgen anti-HIV fusion protein -- Idun Anti-complement ... anti-HIV proteins - Cangene Anti-CTLA-4 MAb Anti-HM1-24 MAb -- Chugai Anti-EGFR catalytic antibody -- Hesed Anti-hR3 MAb Anti-Human-Carcinoma-Antigen MAb -anti-EGFR immunotoxin -- IVAX **Epicyte** Anti-EGFR MAb -- Abgenix Anti-ICAM-1 MAb - Boehringer Ingelheim Anti-EGFR MAb 528 Anti-EGFR MAb KSB 107 -- KS Biomedix . Anti-ICAM-1 MAb 1A-29 -- Pharmacia Anti-ICAM-1 MAb HA58 Anti-EGFR MAb-DM1 conjugate --Anti-ICAM-1 MAb YN1/1.7.4 ImmunoGen Anti-ICAM-3 MAb ICM3 -- ICOS Anti-EGFR MAb-LA1 --Anti-idiotype breast cancer vaccine 11D10 Anti-EGFR sheep MAb -- KS Biomedix Anti-idiotype breast cancer vaccine Anti-FAP MAb F19-I-131 ACA14C5 --Anti-Fas IgM MAb CH11 Anti-idiotype cancer vaccine -- ImClone Anti-Fas MAb Jo2 Systems/Merck KGaA ImClone, Viventia Anti-Fas MAb RK-8 Anti-Flt-1 monoclonal antibodies -- ImClone Anti-idiotype cancer vaccine 1A7 -- Titan Anti-fungal peptides -- State University of Anti-idiotype cancer vaccine 3H1 -- Titan New York Anti-idiotype cancer vaccine TriAb -- Titan antifungal tripeptides -- BTG Anti-ganglioside GD2 antibody-interleukin-2 Anti-idiotype Chlamydia trachomatis vaccine fusion protein -- Lexigen Anti-idiotype colorectal cancer vaccine --Anti-GM2 MAb -- Kyowa Novartis Anti-GM-CSF receptor monoclonal Anti-idiotype colorectal cancer vaccine antibodies -- AMRAD Anti-gp130 MAb -- Tosoh Anti-idiotype melanoma vaccine -- IDEC Anti-HCA monoclonal antibodies --**Pharmaceuticals** AltaRex/Epigen Anti-idiotype ovarian cancer vaccine ACA Anti-hCG antibodies -- Abgenix/AVI BioPharma Anti-idiotype ovarian cancer vaccine AR54 -- AltaRex

### FIG. 28D

Anti-L-selectin monoclonal antibodies --Anti-idiotype ovarian cancer vaccine CA-125 – AltaRex, Biomira Protein Design Labs, Abgenix, Stanford Anti-IgE catalytic antibody -- Hesed Biomed University Anti-MBL monoclonal antibodies --Anti-IgE MAb E26 -- Genentech Alexion/Brigham and Women's Hospital Anti-IGF-1 MAb Anti-MHC monoclonal antibodies anti-inflammatory -- GeneMax Anti-MIF antibody humanised – IDEC, anti-inflammatory peptide -- BTG Cytokine PharmaSciences anti-integrin peptides -- Burnha Anti-interferon-alpha-receptor MAb 64G12 -- Anti-MRSAVRSA sheep MAb -- KS Biomedix Holdings Pharma Pacific Management Anti-interferon-gamma MAb -- Protein Anti-mu MAb -- Novartis Anti-MUC-1 MAb Design Labs Anti-interferon-gamma polyclonal antibody - Anti-MUC 18 Anti-Nogo-A MAb IN1 - Advanced Biotherapy Anti-nuclear autoantibodies -- Procyon Anti-interleukin-10 MAb -Anti-ovarian cancer monoclonal antibodies -Anti-interleukin-12 MAb -Anti-interleukin-1-beta polyclonal antibody -- - Dompe Anti-p185 monoclonal antibodies R&D Systems Anti-p43 MAb Anti-interleukin-2 receptor MAb 2A3 Antiparasitic vaccines Anti-interleukin-2 receptor MAb 33B3-1 -Anti-PDGF/bFGF sheep MAb -- KS Immunotech Biomedix Anti-interleukin-2 receptor MAb ART-18 Anti-properdin monoclonal antibodies --Anti-interleukin-2 receptor MAb LO-Tact-1 Abgenix/Gliatech Anti-interleukin-2 receptor MAb Mikbeta1 Anti-PSMA (prostrate specific membrane Anti-interleukin-2 receptor MAb NDS61 antigen) Anti-interleukin-4 MAb 11B11 Anti-PSMA MAb J591 -- BZL Biologics Anti-interleukin-5 MAb - Wallace Anti-Rev MAb gene therapy -Laboratories Anti-RSV antibodies - Epicyte, Intracell Anti-interleukin-6 MAb - Centocor, Anti-RSV monoclonal antibodies --Diaclone, Pharmadigm Medarex/MedImmune, Applied Molecular Anti-interleukin-8 MAb – Abgenix Anti-interleukin-8 MAb - Xenotech Evolution/Medimmune Anti-RSV MAb, inhalation --Anti-JL1 MAb Aikermes/Medimmune Anti-Klebsiella sheep MAb -- KS Biomedix Anti-RT gene therapy Antisense K-ras RNA gene therapy Anti-Laminin receptor MAb-liposomal Anti-SF-25 MAb doxorubicin conjugate Anti-sperm antibody -- Epicyte Anti-LCG MAb -- Cytoclonal Anti-Tac(Fv)-PE38 conjugate Anti-lipopolysaccharide MAb -- VitaResc Anti-TAPA/CD81 MAb AMP1

FIG. 28E

Anti-tat gene therapy

AOP-RANTES — Senetek Anti-TCR-alphabeta MAb H57-597 Apan-CH - Praecis Pharmaceuticals Anti-TCR-alphabeta MAb R73 APC-8024 - Demegen Anti-tenascin MAb BC-4-I-131 ApoA-1 -- Milano, Pharmacia Anti-TGF-beta human monoclonal Apogen -- Alexion antibodies -- Cambridge Antibody Tech., apolipoprotein A1 - Avanir Genzyme Apolipoprotein E - Bio-Tech. General Anti-TGF-beta MAb 2G7 - Genentech Applaggin - Biogen Antithrombin III - Genzyme Transgenics, aprotinin - ProdiGene Aventis, Bayer, Behringwerke, CSL, APT-070C - AdProTech Myriad AR 177 - Aronex Pharmaceuticals Anti-Thy1 MAb AR 209 - Aronex Pharmaceuticals, Anti-Thy1.1 MAb Anti-tissue factor/factor VIIA sheep MAb -**Antigenics** AR545C KS Biomedix ARGENT gene delivery systems – ARIAD Anti-TNF monoclonal antibodies -Arresten Centocor, Chiron, Peptech, Pharacia, ART-123 - Asahi Kasei Serono Anti-TNF sheep MAb -- KS Biomedix arvisulfatase B -- BioMarin Arvisulfatase B, Recombinant human --**Holdings** BioMarin Anti-TNFalpha MAb - Genzyme Anti-TNFalpha MAb B-C7 - Diaclone AS 1051 — Ajinomoto ASI-BCL - Intracell Anti-tooth decay MAb -- Planet BioTech. Asparaginase - Merck Anti-TRAIL receptor-1 MAb - Takeda ATL-101 - Alizyme Antitumour RNases - NIH Atrial natriuretic peptide - Pharis Anti-VCAM MAb 2A2 - Alexion Aurintricarboxylic acid-high molecular Anti-VCAM MAb 3F4 - Alexion weight Anti-VCAM-1 MAb Autoimmune disorders -- GPC Anti-VEC MAb - ImClone Biotech/MorphoSys Anti-VEGF MAb -- Genentech Autoimmune disorders and transplant Anti-VEGF MAb 2C3 rejection -- Bristol-Myers Squibb/Genzyme Anti-VEGF sheep MAb - KS Biomedix Holdings Autoimmune disorders/cancer -- . Anti-VLA-4 MAb HP1/2 -- Biogen Abgenix/Chiron, CuraGen Anti-VLA-4 MAb PS/2 Autotaxin Anti-VLA-4 MAb R1-2 Avicidin - NeoRx Anti-VLA-4 MAb TA-2 axogenesis factor-1 - Boston Life Sciences Anti-VAP-1 human MAb Axokine – Regeneron Anti-VRE sheep MAb -- KS Biomedix B cell lymphoma vaccine - Biomira Holdings B7-1 gene therapy -ANUP -- TranXenoGen BABS proteins - Chiron ANUP-1 -- Pharis

**FIG. 28F** 

BMP 2 -- Genetics Institute/Medtronic-BAM-002 -- Novelos Therapeutics Sofamor Danek, Genetics Institute/ Basiliximab (anti CD25 MAb) -- Novartis Collagenesis, Genetics Bay-16-9996 -- Bayer Institute/Yamanouch Bay-39-9437 -- Bayer BIVIP 2 gene therapy Bay-50-4798 - Bayer BMP 52 -- Aventis Pasteur, Biopharm BB-10153 -- British Biotech BMP-2 - Genetics Institute BBT-001 -- Bolder BioTech. BMS 182248 -- Bristol-Myers Squibb BBT-002 -- Bolder BioTech. BMS 202448 -- Bristol-Myers Squibb BBT-003 -- Bolder BioTech. bone growth factors - IsoTis BBT-004 -- Bolder BioTech. BPC-15 -- Pfizer BBT-005 -- Bolder BioTech. brain natriuretic peptide -BBT-006 -- Bolder BioTech. Breast cancer -- Oxford BBT-007 -- Bolder BioTech. GlycoSciences/Medarex BCH-2763 -- Shire Breast cancer vaccine -- Therion Biologics, BCSF - Millenium Biologix Oregon BDNF -- Regeneron -- Amgen Becaplermin -- Johnson & Johnson, Chiron BSSL -- PPL Therapeutics BST-2001 - BioStratum Bectumomab -- Immunomedics BST-3002 -- BioStratum Beriplast -- Aventis Beta-adrenergic receptor gene therapy --BTI 322 butyrylcholinesterase -- Shire University of Arkansas C 6822 -- COR Therapeutics bFGF - Scios C1 esterase inhibitor -- Pharming BI 51013 -- Behringwerke AG C3d adjuvant -- AdProTech BIBH 1 -- Boehringer Ingelheim CAB-2.1 - Millennium BIM-23190 -- Beaufour-lpsen calcitonin - Inhale Therapeutics Systems, birch pollen immunotherapy - Pharmacia Aventis, Genetronics, TranXenoGen, bispecific fusion proteins - NIH Unigene, Rhone Poulenc Rohrer Bispecific MAb 2B1 -- Chiron calcitonin -- oral -- Nobex, Emisphere, Bitistatin Pharmaceutical Discovery BIWA 4 -- Boehringer Ingelheim Calcitonin gene-related peptide -- Asahi blood substitute -- Northfield, Baxter Inti. Kasei -- Unigene BLP-25 -- Biomira BLS-0597 -- Boston Life Sciences calcitonin, human -- Suntory calcitonin, nasal -- Novartis, Unigene BLyS -- Human Genome Sciences calcitonin, Panoderm -- Elan BLyS radiolabelled - Human Genome calcitonin, Peptitrol - Shire Sciences calcitonin, salmon - Therapicon BM 06021 -- Boehringer Mannheim calin -- Biopharm BM-202 -- BioMarin Calphobindin I BM-301 -- BioMarin calphobindin I -- Kowa BM-301 -- BioMarin caireticulin -- NYU BM-302 -- BioMarin

FIG. 28G

CD4 fusion toxin — Senetek Campath-1G CD4 IgG -- Genentech Campath-1M CD4 receptor antagonists cancer therapy -- Cangene cancer vaccine - Aixlie, Aventis Pasteur, Pharmacopeia/Progenics CD4 soluble -- Progenics Center of Molecular Immunology, YM CD4, soluble - Genzyme Transgenics BioSciences, Cytos, Genzyme, CD40 ligand - Immunex Transgenics, Globelmmune, Igeneon, CD4-ricin chain A - Genentech ImClone, Virogenetics, InterCell, Iomai, CD59 gene therapy -- Alexion Jenner Biotherapies, Memorial Sloan-Kettering Cancer Center, Sydney Kimmel CD8 TIL cell therapy -- Aventis Pasteur CD8, soluble -- Avidex Cancer Center, Novavax, Protein CD95 ligand - Roche Sciences, Argonex, SIGA CDP 571 - Celitech Cancer vaccine ALVAC-CEA B7.1 --CDP 850 - Celltech Aventis Pasteur/Therion Biologics CDP-860 (PEG-PDGF MAb) -- Celltech Cancer vaccine CEA-TRICOM -- Aventis CDP 870 -- Celltech Pasteur/Therion Biologics CDS-1 -- Emest Orlando Cancer vaccine gene therapy - Cantab Cedelizumab -- Ortho-McNeil **Pharmaceuticals** Cetermin -- Insmed Cancer vaccine HER-2/neu - Corixa CETP vaccine -- Avant Cancer vaccine THERATOPE - Biomira Cetrorelix cancer vaccine, PolyMASC - Valentis Cetuximab Candida vaccine -- Corixa, Inhibitex CGH 400 -- Novartis Canstatin -- ILEX CGP 42934 - Novartis CAP-18 -- Panorama CGP 51901 - Tanox Cardiovascular gene therapy -- Collateral CGRP - Unigene Therapeutics CGS 27913 -- Novartis carperitide - Suntory CGS 32359 - Novartis Casocidin-1 -- Pharis Chagas disease vaccine -- Corixa CAT 152 - Cambridge Antibody Tech. chemokines -- Immune Response CAT 192 - Cambridge Antibody Tech. CHH 380 -- Novartis CAT 213 -- Cambridge Antibody Tech. chitinase - Genzyme, ICOS Catalase— Enzon Chlamydia pneumoniae vaccine -- Antex Cat-PAD - Circassia **Biologics** CB 0006 - Celltech Chlamydia trachomatis vaccine -- Antex CCK(27-32)-- Akzo Nobel **Biologics** CCR2-641 -- NIH Chlamydia vaccine - GlaxoSmithKline CD, Procept -- Paligent Cholera vaccine CVD 103-HgR -- Swiss CD154 gene therapy Serum and Vaccine Institute Beme CD39 -- Immunex Cholera vaccine CVD 112 -- Swiss Serum CD39-L2 -- Hyseq and Vaccine Institute Berne CD39-L4 -- Hyseq

FIG. 28H

CRL 1605 - CytRx Cholera vaccine inactivated oral - SBL CS-560 - Sankyo Vaccin CSF - ZymoGenetics Chrysalin -- Chrysalis BioTech. CSF-G - Hangzhou, Dong-A, Hanmi CI-782 -- Hitachi Kase CSF-GM - Cangene, Hunan, LG Chem Ciliary neurotrophic factor - Fidia, Roche CSF-M - Zarix CIM project -- Active Biotech CT 1579 -- Merck Frosst CL 329753 - Wyeth-Ayerst CT 1786 - Merck Frosst CL22, Cobra -- ML Laboratories CT-112<sup>^</sup> -- BTG Clenoliximab -- IDEC CTB-134L - Xenova Clostridium difficile antibodies -- Epicyte CTC-111 - Kaketsuken clotting factors -- Octagene CTGF - FibroGen CMB 401 -- Celltech CTLA4-Ig -- Bristol-Myers Squibb CNTF -- Sigma-Tau CTLA4-lg gene therapy --Cocaine abuse vaccine - Cantab, CTP-37 -- AVI BioPharma ImmuLogic, Scripps C-type natriuretic peptide — Suntory coccidiomycosis vaccine -- Arizo CVS 995 - Corvas Intl. collagen - Type I - Pharming CX 397 – Nikko Kyodo Collagen formation inhibitors - FibroGen Collagen/hydroxyapatite/bone growth factor CY 1747 -- Epimmune CY 1748 -- Epimmune - Aventis Pasteur, Biopharm, Orquest Cyanovirin-N collagenase -- BioSpecifics Colorectal cancer vaccine -- Wistar Institute Cystic fibrosis therapy -- CBR/IVAX CYT 351 Component B, Recombinant -- Serono Connective tissue growth factor inhibitors -- cytokine Traps -- Regeneron cytokines - Enzon, Cytoclonal FibroGen/Taisho Cytomegalovirus glycoprotein vaccine --Contortrostatin Chiron, Aquila Biopharmaceuticals, contraceptive vaccine -- Zonagen Aventis Pasteur, Virogenetics Contraceptive vaccine hCG Cytomegalovirus vaccine live -- Aventis Contraceptive vaccine male reversible --Pasteur IMMUCON Cytosine deaminase gene therapy -Contraceptive vaccine zona pellucida --GlaxoSmithKline DA-3003 -- Dong-A Copper-64 labelled MAb TETA-1A3 -- NCI DAB389interleukin-6 -- Senetek Coralyne DAB389interleukin-7 Corsevin M Daclizumab (anti-IL2R MAb) - Protein C-peptide analogues -- Schwarz CPI-1500 -- Consensus Design Labs DAMP<sup>^</sup> – Incyte Genomics CRF — Neurobiological Tech. Daniplestim -- Pharmacia cRGDfV pentapeptide -darbepoetin alfa -- Amgen CRL 1095 -- CytRx DBI-3019 -- Diabetogen CRL 1336 -- CytRx

FIG. 281

Duteplase -- Baxter Intl. DCC -- Genzyme DWP-401 - Daewoong DDF -- Hyseq DWP-404 -- Daewoong decorin ~ Integra, Telios defensins - Large Scale Biology DWP-408 -- Daewoong Dx 88 (Epi-KAL2) -- Dyax DEGR-VIIa Dx 890 (elastin inhibitors) - Dyax Delmmunised antibody 3B6/22 AGEN E coli O157 vaccine -- NIH Deimmunised anti-cancer antibodies --E21-R -- BresaGen Biovation/Viragen Eastern equine encephalitis virus vaccine -Dendroamide A Dengue vaccine -- Bavarian Nordic, Merck Echicetin --Echinhibin 1 denileukin diftitox -- Ligand Echistatin – Merck DES-1101 - Desmos Echitamine desirudin -- Novartis Ecromeximab – Kyowa Hakko desmopressin -- Unigene Desmoteplase - Merck, Schering AG EC-SOD -- PPL Therapeutics Eculizumab (5G1.1) -- Alexion Destabilase Diabetes gene therapy - DeveloGen, Pfizer EDF - Ajinomoto EDN derivative - NIH Diabetes therapy -- Crucell Diabetes type 1 vaccine - Diamyd EDNA -- NIH Edobacomab -- XOMA **Therapeutics** Edrecolomab -- Centocor DiaCIM -- YM BioSciences EF 5077 dialytic oligopeptides -- Research Corp Efalizumab - Genentech Diamyd -- Diamyd Therapeutics EGF fusion toxin - Seragen, Ligand DiaPep227-- Pepgen EGF-P64k vaccine -- Center of Molecular DiavaX -- Corixa Immunology Digoxin MAb -- Glaxo EL 246 - LigoCyte Diphtheria tetanus pertussis-hepatitis B elastase inhibitor - Synergen vaccine -- GlaxoSmithKline elcatonin -- Therapicon DIR therapy — Solis Therapeutics — EMD 72000 -- Merck KGaA DNase -- Genentech Emdogain -- BIORA Dornase alfa -- Genentech emfilermin - AMRAD Dornase alfa, inhalation -- Genentech Emoctakin - Novartis Doxorubicin-anti-CEA MAb conjugate enamel matrix protein -- BIORA **Immunomedics** Endo III -- NYU DP-107 -- Trimeris endostatin -- EntreMed, Pharis drotrecogin alfa -- Eli Lilly Enhancins -- Micrologix DTctGMCSF Enlimomab - Isis Pharm. DTP-polio vaccine -- Aventis Pasteur Enoxaparin sodium -- Pharmuka DU 257-KM231 antibody conjugate -enzyme linked antibody nutrient depletion Kvowa therapy -- KS Biomedix Holdings

**FIG. 28J** 

dural graft matrix -- Integra

Factor VII -- Novo Nordisk, Bayer, Baxter Eosinophil-derived neutralizing agent -EP-51216 -- Asta Medica Intl. Factor VIIa -- PPL Therapeutics, EP-51389 -- Asta Medica EPH family ligands - Regeneron ZvmoGenetics Epidermal growth factor - Hitachi Kasei, Factor VIII - Bayer Genentech, Beaufour-Ipsen, CLB, Inex, Octagen, Pharmacia, Johnson & Johnson Epidermal growth factor fusion toxin— Pharming Factor VIII -- PEGylated -- Bayer Senetek Factor VIII fragments - Pharmacia Epidermal growth factor-genistein – Factor VIII gene therapy -- Targeted EPI-HNE-4 -- Dvax Genetics EPI-KAL2 -- Dyax Factor VIII sucrose formulation - Bayer, Epoetin-alfa -- Amgen, Dragon Genentech Pharmaceuticals, Nanjing Huaxin Factor VIII-2 -- Bayer Epratuzumab -- Immunomedics Factor VIII-3 -- Bayer Epstein-Barr virus vaccine --Factor Xa inhibitors - Merck, Novo Nordisk, Aviron/SmithKline Beecham, Bioresearch Mochida Eptacog alfa -- Novo Nordisk Factor XIII -- ZymoGenetics Eptifibatide -- COR Therapeutics Factors VIII and IX gene therapy -- Genetics erb-38 --Institute/Targeted Genetics Erlizumab -- Genentech erythropoletin -- Alkermes, ProLease, Dong-Famoxin -- Genset Fas (delta) TM protein – LXR BioTech. A, Elanex, Genetics Institute, LG Chem, Fas TR -- Human Genome Sciences Protein Sciences, Serono, Snow Brand, SRC VB VECTOR, Transkaryotic Felvizumab -- Scotgen FFR-VIIa - Novo Nordisk Therapies FG-001 -- F-Gene Erythropoietin Beta -- Hoffman La Roche FG-002 - F-Gene Erythropoietin/Epoetin alfa -- Chugai FG-004 - F-Gene Escherichia coli vaccine - North American FG-005 - F-Gene Vaccine, SBL Vaccin, Swiss Serum and FGF + fibrin -- Repair Vaccine Institute Berne Fibrimage -- Bio-Tech. General etanercept -- Immunex fibrin-binding peptides – ISIS Innovation examorelin – Mediolanum fibringen – PPL Therapeutics, Pharming Exendin 4 -- Amylin fibroblast growth factor - Chiron, NYU, exonuclease VII Ramot, ZymoGenetics F 105 -- Centocor fibrolase conjugate - Schering AG F-992 -- Fornix Filgrastim -- Amgen Factor IX -- Alpha Therapeutics, Welfide filgrastim - PDA modified - Xencor Corp., CSL, enetics institute/AHP, FLT-3 ligand -- Immunex Pharmacia, PPL Therapeutics FN18 CRM9 -Factor IX gene therapy -- Cell Genesys

#### FIG. 28K

glutamate decarboxylase -- Genzyme follistatin -- Biotech Australia, Human Transgenics Therapeutics Glycoprotein S3 - Kureha follitropin alfa - Alkermes, ProLease, GM-CSF - Immunex PowderJect, Serono, Akzo Nobel GM-CSF tumour vaccine - PowderJect Follitropin Beta – Bayer, Organon GnRH immunotherapeutic -- Protherics FP 59 Goserelin (LhRH antagonist) -- AstraZeneca FSH -- Ferring gp75 antigen -- ImClone FSH + LH - Ferring gp96 - Antigenics F-spondin -- CeNeS GPI 0100 - Galenica fusion protein delivery system -- UAB GR 4991W93 - GlaxoSmithKline Research Foundation Granulocyte colony-stimulating factor -fusion toxins -- Boston Life Sciences Dong-A G 5598 -- Genentech Granulocyte colony-stimulating factor GA-II -- Transkaryotic Therapies coniugate Gamma-interferon analogues -- SRC VB grass allergy therapy -- Dynavax **VECTOR** GRF1-44 -- ICN Ganirelix -- Roche Growth Factor – Chiron, Atrigel, Atrix, gastric lipase -- Meristem Innogenetics, ZymoGenetics, Novo Gavilimomab growth factor peptides -- Biotherapeutics G-CSF – Amgen, SRC VB VECTOR growth hormone - LG Chem GDF-1 -- CeNeS growth hormone, Recombinant human -GDF-5 - Biopharm Serono GDNF (glial derived neurotrophic factor) --GT 4086 -- Gliatech Amgen GW 353430 -- GlaxoSmithKline gelsolin - Biogen GW-278884 - GlaxoSmithKline Gemtuzumab ozogamicin -- Celltech H 11 - Viventia Biotech Gene-activated epoetin-alfa -- Aventis H5N1 influenza A virus vaccine - Protein Pharma - Transkaryotic Therapies Sciences Glanzmann thrombasthenia gene therapy haemoglobin -- Biopure Glatiramer acetate - Yeda haemoglobin 3011, Recombinant -- Baxter glial growth factor 2 - CeNeS GLP-1 - Amylin, Suntory, TheraTech, Healthcare haemoglobin crosfumaril – Baxter Intl. Watson haemoglobin stabilized - Ajinomoto GLP-1 peptide analogues -- Zealand haemoglobin, recombinant - Apex **Pharaceuticals** HAF - Immune Response glucagon -- Eli Lilly, ZymoGenetics Glucagon-like peptide-1 7-36 amide --Hantavirus vaccine HB 19 Suntory Glucogen-like peptide -- Amylin HBNF -- Regeneron HCC-1 - Pharis Glucocerebrosidase - Genzyme hCG - Milkhaus

#### FIG. 28L

hCG vaccine -- Zonagen HE-317 -- Hollis-Eden Pharmaceuticals Heat shock protein cancer and influenza Takeda vaccines -- StressGen Helicobacter pylori vaccine - Acambis, AstraZeneca/CSL, Chiron, Provalis Helistat-G - GalaGen Morley Hemolink – Hemosol hepapoietin -- Snow Brand heparanase -- InSight heparinase I -- Ibex heparinase III -- Ibex Hepatitis A vaccine -- American Biogenetic HIC 1 Sciences Hepatitis A vaccine inactivated Hepatitis A vaccine Nothav -- Chiron Hepatitis A-hepatitis B vaccine --GlaxoSmithKline hepatitis B therapy -- Tripep Hepatitis B vaccine - Amgen, Chiron SpA, Meiji Milk, NIS, Prodeva, PowderJect, VaxGen Rhein Biotech Hepatitis B vaccine recombinant - Evans Vaccines, Epitec Combiotech, Genentech, HIV gp160 DNA vaccine - PowderJect, Medimmune, Merck Sharp & Dohme, Rhein Biotech, Shantha Biotechnics, Vector, Yeda Hepatitis B vaccine recombinant TGP 943 -- HIV HGP-30W vaccine -- CEL-SCI Takeda Hepatitis C vaccine -- Bavarian Nordic, Chiron, Innogenetics Acambis, Hepatitis D vaccine -- Chiron Vaccines Hepatitis E vaccine recombinant --Genelabs/GlaxoSmithKline, Novavax hepatocyte growth factor - Panorama, Sosei hepatocyte growth factor kringle fragments - EntreMed Her-2/Neu peptides -- Corixa

Herpes simplex glycoprotein DNA vaccine -Merck, Wyeth-Lederle Vaccines-Malvern, Genentech, GlaxoSmithKline, Chiron, Herpes simplex vaccine -- Cantab Pharmaceuticals, CEL-SCI, Henderson Herpes simplex vaccine live -- ImClone Systems/Wyeth-Lederle, Aventis Pasteur HGF derivatives -- Dompe hIAPP vaccine -- Crucell Hib-hepatitis B vaccine -- Aventis Pasteur HIP-- Altachem Hirudins - Biopharma, Cangene, Dongkook, Japan Energy Corporation, Pharmacia Corporation, SIR International, Sanofi-Synthelabo, Sotragene, Rhein Biotech HIV edible vaccine -- ProdiGene HIV gp120 vaccine - Chiron, Ajinomoto, GlaxoSmithKline, ID Vaccine, Progenics, HIV gp120 vaccine gene therapy -Aventis Pasteur, Oncogen, Hyland Immuno, Protein Sciences HIV gp41 vaccine -- Panacos · HIV immune globulin -- Abbott, Chiron HIV peptides - American Home Products HIV vaccine - Applied bioTech., Axis Genetics, Biogen, Bristol-Myers Squibb, · Genentech, Korea Green Cross, NIS, Oncogen, Protein Sciences Corporation, Terumo, Tonen Corporation, Wyeth-Averst, Wyeth-Lederle Vaccines-Malvern, Advanced BioScience Laboratories, Bavarian Nordic, Bavarian Nordic/Statens Serum Institute, GeneCure, Immune Response, Progenics, Therion Biologics, United Biomedical, Chiron

#### **FIG. 28M**

HIV vaccine vCP1433 -- Aventis Pasteur Human monoclonal antibodies --Medarex/Northwest Biotherapeutics, HIV vaccine vCP1452 -- Aventis Pasteur HIV vaccine vCP205 -- Aventis Pasteur Medarex/Seattle Genetics human netrin-1 -- Exelixis HL-9 -- American BioScience human papillomavirus antibodies -- Epicyte HM-9239 -- Cytran Human papillomavirus vaccine -- Biotech HML-103 -- Hemosol Australia, IDEC, StressGen HML-104 -- Hemosol Human papillomavirus vaccine MEDI 501 --HML-105 -- Hemosol MedImmune/GlaxoSmithKline HML-109 -- Hemosol Human papillomavirus vaccine MEDI HML-110 -- Hemosol 503/MEDI 504 --HML-121 -- Hemosol Medlmmune/GlaxoSmithKline hNLP -- Pharis Human papillomavirus vaccine TA-CIN --Hookworm vaccine Cantab Pharmaceuticals host-vector vaccines -- Henogen Human papillomavirus vaccine TA-HPV --HPM 1 -- Chugai Cantab Pharmaceuticals HPV vaccine -- MediGene Human papillomavirus vaccine TH-GW ---HSA -- Meristem Cantab/GlaxoSmithKline HSF -- StressGen human polyclonal antibodies - Biosite/Eos HSP carriers -- Weizmann, Yeda, Peptor BioTech./ Medarex HSPPC-70 -- Antigenics HSPPC-96, pathogen-derived -- Antigenics human type II anti factor VIII monoclonal antibodies -- ThromboGenics HSV 863 -- Novartis humanised anti glycoprotein lb murine HTLV-I DNA vaccine monoclonal antibodies -- ThromboGenics HTLV-I vaccine HumaRAD -- Intracell HTLV-II vaccine -- Access HuMax EGFR -- Genmab HU 901 -- Tanox HuMax-CD4 -- Medarex Hu23F2G - ICOS HuMax-IL15 -- Genmab HuHMFG1 HYB 190 -- Hybridon HumaLYM -- Intracell HYB 676 -- Hybridon Human krebs statika - Yamanouchi I-125 MAb A33 -- Celltech human monoclonal antibodies --Ibritumomab tiuxetan -- IDEC Abgenix/Biogen, Abgenix/ Corixa, Abgenix/Immunex, Abgenix/Lexicon, IBT-9401 -- Ibex IBT-9402 -- Ibex Abgenix/ Pfizer, Athersys/Medarex, IC 14 - ICOS Biogen/MorphoSys, CAT/Searle, Idarubicin anti-Ly-2.1 --Centocor/Medarex, Corixa/Kirin Brewery, IDEC 114 -- IDEC Corixa/Medarex, Eos BioTech./Medarex, IDEC 131 - IDEC Eos/Xenerex, Exelixis/Protein Design Labs, ImmunoGen/ Raven, Medarex/ IDEC 152 - IDEC B.Twelve, MorphoSys/ImmunoGen, XTL IDM 1 -- IDM IDPS -- Hollis-Eden Pharmaceuticals Biopharmaceuticals/Dyax,

**FIG. 28N** 

iduronate-2-sulfatase -- Transkaryotic insulin -- Autolmmune, Altea, Biobras, BioSante, Bio-Tech. General, Chong Kun Therapies Dang, Emisphere, Flamel, Provalis, Rhein IGF/IBP-2-13 -- Pharis IGN-101 -- Igeneon Biotech, TranXenoGen IK HIR02 -- Iketon insulin (bovine) -- Novartis insulin analogue -- Eli Lilly IL-11 — Genetics Institute/AHP Insulin Aspart -- Novo Nordisk IL-13-PE38 -- NeoPharm insulin detemir -- Novo Nordisk IL-17 receptor -- Immunex IL-18BP - Yeda insulin glargine -- Aventis insulin inhaled - Inhale Therapeutics IL-1Hy1 -- Hyseq Systems, Alkermes IL-1ß -- Celltech insulin oral -- Inovax IL-1ß adjuvant -- Celltech IL-2 -- Chiron insulin, AeroDose -- AeroGen insulin, AERx -- Aradigm IL-2 + IL-12 -- Hoffman La-Roche insulin, BEODAS -- Elan IL-6/sIL-6R fusion -- Hadasit insulin, Biphasix -- Helix IL-6R derivative - Tosoh IL-7-Dap 389 fusion toxin -- Ligand insulin, buccal -- Generex insulin, I2R -- Flemington IM-862 -- Cytran insulin, intranasal -- Bentley IMC-1C11 -- ImClone insulin, oral -- Nobex, Unigene imiglucerase -- Genzyme insulin, Orasome -- Endorex Immune globulin intravenous (human) --insulin, ProMaxx -- Epic Hoffman La Roche immune privilege factor - Proneuron insulin, Quadrant -- Elan insulin, recombinant -- Aventis Immunocal -- Immunotec insulin, Spiros -- Elan Immunogene therapy -- Briana Bio-Tech Immunoliposomal 5-fluorodeoxyuridineinsulin, Transfersome -- IDEA insulin, Zymo, recombinant -- Novo Nordisk dipalmitate -immunosuppressant vaccine -- Aixlie insulinotropin -- Scios Insulysin gene therapy immunotoxin -- Antisoma, NIH ImmuRAIT-Re-188 – Immunomedics integrin antagonists -- Merck interferon (Alpha2) -- SRC VB VECTOR, imreg-1 -- Imreg Viragen, Dong-A, Hoffman La-Roche, infertility -- Johnson & Johnson, E-TRANS Genentech Infliximab -- Centocor Influenza virus vaccine -- Aventis Pasteur, interferon - BioMedicines, Human Genome Sciences Protein Sciences inhibin -- Biotech Australia, Human interferon (Alfa-n3)---Interferon Sciences Therapeutics Inhibitory G protein gene therapy interferon (Alpha), Biphasix -- Helix INKP-2001 -- InKine Inolimomab -- Diaclone

FIG. 280

IL-2/ diphtheria toxin - Ligand interferon (Alpha)—Amgen, BioNative, Interleukin-3 - Cangene Novartis, Genzyme Transgenics, Interleukin-4 - Immunology Ventures, Hayashibara, Inhale Therapeutics Sanofi Winthrop, Schering-Plough, Systems, Medusa, Flamel, Dong-A, Immunex/ Sanofi Winthrop, Bayer, Ono GeneTrol, Nastech, Shantha, interleukin-4 + TNF-Alpha -- NIH Wassermann, LG Chem, Sumitomo, interleukin-4 agonist -- Bayer Aventis, Behring EGIS, Pepgen, Servier, interleukin-4 fusion toxin - Ligand Rhein Biotech, Interleukin-4 receptor - Immunex, Immun interferon (Alpha2A) Interleukin-6 - Ajinomoto, Cangene, Yeda, interferon (Alpha2B) - Enzon, Schering-Genetics Institute, Novartis Plough, Biogen, IDEA interferon (Alpha-N1) -- GlaxoSmithKline interleukin-6 fusion protein interleukin-6 fusion toxin - Ligand, Serono interferon (beta) - Rentschler, GeneTrol, interleukin-7 -- IC Innovations Meristem, Rhein Biotech, Toray, Yeda, interleukin-7 receptor -- Immunex Daiichi, Mochida interleukin-8 antagonists -- Kyowa interferon (Beta1A) - Serono, Biogen Hakko/Millennium/Pfizer interferon (beta1A), inhale -- Biogen interleukin-9 antagonists -- Genaera interferon (ß1b)-- Chiron Interleukin-10 - DNAX, Schering-Plough interferon (tau) - Pepgen: Interleukin-10 gene therapy -Interferon alfacon-1 - Amgen interleukin-12 -- Genetics Institute, Hoffman Interferon alpha-2a vaccine La-Roche Interferon Beta 1b - Schering/Chiron, interleukin-13 -- Sanofi interMune Interferon Gamma -- Boehringer Ingelheim, interleukin-13 antagonists -- AMRAD interleukin-13-PE38QQR Sheffield, Rentschler, Hayashibara interleukin-15 - Immunex interferon receptor, Type I - Serono interferon(Gamma1B) - Genentech interleukin-16 - Research Corp interleukin-18 - GlaxoSmithKline Interferon-alpha-2b + ribavirin - Biogen, Interleukin-18 binding protein -- Serono ICN lor-P3 -- Center of Molecular immunology interferon-alpha-2b gene therapy --IP-10 -- NIH Schering-Plough IPF -- Metabolex Interferon-con1 gene therapy -IR-501 -- Immune Response interleukin-1 antagonists - Dompe ISIS 9125 - Isis Pharmaceuticals Interleukin-1 receptor antagonist - Abbott ISURF No. 1554 - Millennium Bioresearch, Pharmacia ISURF No. 1866 - Iowa State Univer. Interleukin-1 receptor type I — Immunex ITF-1697 -- Italfarmaco interleukin-1 receptor Type II -- Immunex IxC 162 -- Ixion Interleukin-1 trap -- Regeneron Interleukin-1-alpha -- Immunex/Roche J 695 - Cambridge Antibody Tech., Genetics Inst., Knoli interleukin-2 - SRC VB VECTOR, Jagged + FGF -- Repair Ajinomoto, Biomira, Chiron

FIG. 28P

JKC-362 -- Phoenix Pharmaceuticals leptin, 2nd-generation - Amgen leridistim - Pharmacia JTP-2942 – Japan Tobacce Juman monoclonal antibodies -leuprolide, ProMaxx -- Epic leuprorelin, oral -- Unigene Medarex/Raven LeuTech -- Papatin K02 -- Axys Pharmaceuticals LEX 032 -- SuperGen Keliximab -- IDEC LiDEPT -- Novartis Keyhole limpet haemocyanin KGF -- Amgen Lintuzumab (anti-CD33 MAb) -- Protein KM 871 -- Kyowa Design Labs lipase - Altus Biologics KPI 135 -- Scios lipid A vaccine -- EntreMed KPI-022 -- Scios lipid-linked anchor Tech. - ICRT, ID Kringle 5 Biomedical KSB 304 liposome-CD4 Tech. -- Sheffield KSB-201 -- KS Biomedix Listeria monocytogenes vaccine L 696418 -- Merck . LMB 1 L 703801 -- Merck LMB 7 L1 -- Acorda LMB 9 -- Battelle Memorial Institute, NIH L-761191 -- Merck lactoferrin – Meristem, Pharming, Agennix LM-CD45 -- Cantab Pharmaceuticals lactoferrin cardio - Pharming lovastatin -- Merck LSA-3 LAG-3 -- Serono LT-ß receptor -- Biogen LAIT - GEMMA lung cancer vaccine -- Corixa LAK cell cytotoxin -- Arizona lamellarins - PharmaMar/University of lusupultide -- Scios L-Vax -- AVAX Malaga laminin A peptides -- NIH LY 355455 - Eli Lilly lanoteplase -- Genetics Institute LY 366405 -- Eli Lilly laronidase -- BioMarin LY-355101 -- Eli Lilly Lyme disease DNA vaccine -- Vical/Aventis Lassa fever vaccine Pasteur LCAT -- NIH LDP 01 -- Millennium Lyme disease vaccine -- Aquila LDP 02 -- Millennium Biopharmaceuticals, Aventis, Pasteur, Symbicom, GlaxoSmithKline, Hyland Lecithinized superoxide dismutase --Immuno, Medimmune Seikagaku LeIF adjuvant -- Corixa Lymphocytic choriomeningitis virus vaccine lymphoma vaccine - Biomira, Genitope leishmaniasis vaccine -- Corixa LYP18 lenercept -- Hoffman La-Roche lys plasminogen, recombinant Lenograstim -- Aventis, Chugai Lysosomal storage disease gene therapy -lepirudin -- Aventis leptin - Amgen, IC Innovations Avigen Leptin gene therapy -- Chiron Corporation Iysostaphin -- Nutrition 21

**FIG. 28Q** 

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MEDI 507 - BioTransplant M 23 -- Gruenenthal M1 monoclonal antibodies -- Acorda melanin concentrating hormone --Neurocrine Biosciences Therapeutics MA 16N7C2 - Corvas Intl. melanocortins - OMRF Melanoma monoclonal antibodies -- Viragen malaria vaccine -- GlaxoSmithKline, melanoma vaccine -- GlaxoSmithKline, AdProTech, Antigenics, Apovia, Aventis Akzo Nobel, Avant, Aventis Pasteur. Pasteur, Axis Genetics, Behringwerke, Bavarian Nordic, Biovector, CancerVax, CDCP, Chiron Vaccines, Genzyme Genzyme Molecular Oncology, Humbolt, Transgenics, Hawaii, Medimmune, NIH, NYU, Oxxon, Roche/Saramane, Biotech ImClone Systems, Memorial, NYU, Oxxon Australia, Rx Tech Melanoma vaccine Magevac -- Therion Malaria vaccine CDC/NIIMALVAC-1 memory enhancers -- Scios meningococcal B vaccine -- Chiron malaria vaccine.multicomponent meningococcal vaccine -- CAMR mammaglobin -- Corixa Meningococcal vaccine group B conjugate mammastatin -- Biotherapeutics - North American Vaccine mannan-binding lectin -- Natlmmu mannan-MUC1 -- Psiron Meningococcal vaccine group B recombinant -- BioChem Vaccines, **MAP 30** Microscience Marinovir -- Phytera Meningococcal vaccine group Y conjugate -MARstem -- Maret - North American Vaccine MB-015 - Mochida Meningococcal vaccine groups A B and C MBP -- ImmuLogic conjugate -- North American Vaccine MCI-028 -- Mitsubishi-Tokyo Mepolizumab - GlaxoSmithKline MCIF - Human Genome Sciences MDC - Advanced BioScience - Akzo Metastatin – EntreMed, Takeda Met-CkB7 -- Human Genome Sciences Nobel, ICOS met-enkephalin - TNI MDX 11 -- Medarex METH-1 -- Human Genome Sciences MDX 210 -- Medarex methioninase - AntiCancer MDX 22 -- Medarex Methionine lyase gene therapy --MDX 22 MDX 240 -- Medarex AntiCancer Met-RANTES - Genexa Biomedical, MDX 33 MDX 44 – Medarex Serono Metreleptin MDX 447 -- Medarex Microtubule inhibitor MAb MDX H210 - Medarex MDX RA -- Houston BioTech., Medarex Immunogen/Abgenix MGDF - Kirin ME-104 - Pharmexa MGV - Progenics Measles vaccine micrin -- Endocrine Mecasermin -- Cephalon/Chiron, Chiron microplasmin -- ThromboGenics MEDI 488 -- Medimmune MIF -- Genetics Institute MEDI 500

FIG. 28R

MAb 45-2D9- - haematoporphyrin migration inhibitory factor -- NIH Mim CD4.1 - Xycte Therapies conjugate mirostipen -- Human Genome Sciences MAb 4B4 Mitumomab (BEC-2) - ImClone Systems, MAb 4E3-CPA conjugate - BCM Oncologia MAb 4E3-daunorubicin conjugate Merck KGaA MAb 50-6 MK 852 - Merck MAb 50-61A – Institut Pasteur MLN 1202 (Anti-CCR2 monoclonal antibody) - Millenium Pharmaceuticals MAb 5A8 -- Biogen MAb 791T/36-methotrexate conjugate Mobenakin - NIS molgramostim - Genetics Institute, Novartis MAb 7c11.e8 monoclonal antibodies -- Abgenix/Celltech, MAb 7E11 C5-selenocystamine conjugate Immusol/ Medarex, Viragen/ Roslin MAb 93KA9 -- Novartis Institute, Cambridge Antibody Tech./Elan MAb A5B7-cisplatin conjugate --Biodynamics Research, Pharmacla MAb 108 --MAb A5B7-I-131 MAb 10D5 --MAb 14.18-interleukin-2 immunocytokine - MAb A7 MAb A717 -- Exocell Lexigen MAb A7-zinostatin conjugate MAb 14G2a -MAb ABX-RB2 -- Abgenix MAb 15A10 -MAb, ACA 11 MAb 170 -- Biomira MAb AFP-I-131 - Immunomedics MAb 177Lu CC49 --MAb AP1 MAb 17F9 MAb AZ1 MAb 1D7 MAb B3-LysPE40 conjugate MAb 1F7 – Immune Network MAb B4 – United Biomedical MAb 1H10-doxorubicin conjugate MAb B43 Genistein-conjugate MAb 26-2F MAb B43.13-Tc-99m -- Biomira MAb 2A11 MAb B43-PAP conjugate MAb 2E1 -- RW Johnson MAb B4G7-gelonin conjugate MAb 2F5 MAb BCM 43-daunorubicin conjugate -MAb 31.1 — International BioImmune BCM Oncologia Systems MAb BIS-1 MAb 32 - Cambridge Antibody Tech., MAb BMS 181170 -- Bristol-Myers Squibb Peptech MAb BR55-2 MAb 323A3 - Centocor MAb BW494 MAb 3C5 MAb C 242-DM1 conjugate -- ImmunoGen MAb 3F12 MAb C242-PE conjugate MAb 3F8 MAb c30-6 MAb 42/6 MAb CA208-cytorhodin-S conjugate --MAb 425 - Merck KGaA Hoechst Japan MAb 447-52D - Merck Sharp & Dohme MAb CC49 -- Enzon

FIG. 28S

MAb LL2-I-131 — Immunomedics MAb ch14.18 --MAb CH14.18-GM-CSF fusion protein --MAb LL2-Y-90 MAb LS2D617 -- Hybritech Lexigen MAb chCE7 MAb LYM-1-gelonin conjugate MAb CI-137 -- AMRAD MAb LYM-1-I-131 . MAb LYM-1-Y-90 MAb cisplatin conjugate MAb CLB-CD19 MAb LYM-2 -- Peregrine MAb CLB-CD19v MAb M195 MAb M195-bismuth 213 conjugate — MAb CLL-1 -- Peregrine MAb CLL-1-GM-CSF conjugate Protein Design Labs MAb CLL-1-IL-2 conjugate -- Peregrine MAb M195-gelonin conjugate MAb CLN IgG -- doxorubicin conjugates MAb M195-I-131 MAb M195-Y-90-MAb conjugates – Tanox MAb MA 33H1 - Sanofi MAb D612 MAb Dal B02 MAb MAD11 MAb DC101 - ImClone MAb MGb2 MAb MINT5 MAb EA 1 --MAb EC708 -- Biovation MAb MK2-23 MAb EP-5C7 -- Protein Design Labs MAb MOC31 ETA(252-613) conjugate MAb MOC-31-In-111 MAb ERIC-1 -- ICRT MAb MOC-31-PE conjugate MAb F105 gene therapy MAb FC 2.15 MAb MR6 --MAb MRK-16 - Aventis Pasteur MAb G250 -- Centocor MAb GA6 MAb MS11G6 MAb MX-DTPA BrE-3 MAb GA733 MAb Gliomab-H -- Viventia Biotech MAb MY9 MAb Nd2 - Tosoh MAb HB2-saporin conjugate MAb NG-1 -- Hygeia MAb HD 37 -MAb HD37-ricin chain-A conjugate MAb NM01 - Nissin Food **MAb OC 125** MAb HNK20 -- Acambis MAb OC 125-CMA conjugate MAb huN901-DM1 conjugate --ImmunoGen MAb OKI-1 -- Ortho-McNeil MAb OX52 -- Bioproducts for Science MAb I-131 CC49 -- Corixa MAb PMA5 MAb ICO25 MAb PR1 MAb ICR12-CPG2 conjugate MAb prost 30 MAb ICR-62 MAb IRac-ricin A conjugate MAb R-24 MAb K1 MAb R-24 a Human GD3 -- Celltech MAb RFB4-ricin chain A conjugate MAb KS1-4-methotrexate conjugate MAb L6 -- Bristol-Myers Squibb, Oncogen MAb RFT5-ricin chain A conjugate MAb LiCO 16-88 MAb SC 1

#### FIG. 28T

Muc-1 vaccine -- Corixa MAb SM-3 -- ICRT MAb SMART 1D10 -- Protein Design Labs mucosal tolerance -- Aberdeen mullerian inhibiting subst MAb SMART ABL 364 - Novartis muplestim - Genetics Institute, Novartis, MAb SN6f MAb SN6f-deglycosylated ricin A chain DSM Anti-Infectives murine MAb -- KS Biomedix conjugate -Mutant somatropin -- JCR Pharmaceutical MAb SN6i MV 833 -- Toagosei MAb SN7-ricin chain A conjugate Mycoplasma pulmonis vaccine MAb T101-Y-90 conjugate -- Hybritech Mycoprex -- XOMA MAb T-88 -- Chiron myeloperoxidase - Henogen MAb TB94 -- Cancer ImmunoBiology myostatin -- Genetics Institute MAb TEC 11 Nacolomab tafenatox -- Pharmacia MAb TES-23 -- Chugai Nagrecor -- Scios MAb TM31 -- Avant nagrestipen -- British Biotech MAb TNT-1 -- Cambridge Antibody Tech., NAP-5 - Corvas Intl. Peregrine NAPc2 - Corvas Intl. MAb TNT-3 nartograstim -- Kyowa MAb TNT-3 -- IL2 fusion protein --Natalizumab -- Protein Design Labs MAb TP3-At-211 Nateplase – NIH, Nihon Schering MAb TP3-PAP conjugate nateplase -- Schering AG MAb UJ13A -- ICRT NBI-3001 -- Neurocrine Biosci. MAb UN3 NBI-5788 - Neurocrine Biosci. MAb ZME-018-gelonin conjugate NBI-6024 - Neurocrine Biosci. MAb-BC2 - GlaxoSmithKline Nef inhibitors -- BRI MAb-DM1 conjugate -- ImmunoGen Neisseria gonorrhoea vaccine -- Antex MAb-ricin-chain-A conjugate -- XOMA Biologics MAb-temoporfin conjugates Neomycin B-arginine conjugate Monopharm C -- Viventia Biotech Nerelimomab -- Chiron monteplase -- Eisai Nerve growth factor - Amgen - Chiron, montirelin hydrate - Gruenenthal Genentech moroctocog alfa - Genetics Institute Nerve growth factor gene therapy Moroctocog-alfa -- Pharmacia nesiritide citrate -- Scios MP 4 neuregulin-2 -- CeNeS MP-121 -- Biopharm neurocan -- NYU MP-52 -- Biopharm neuronal delivery system -- CAMR MRA -- Chugai Neurophil inhibitory Factor -- Corvas MS 28168 -- Mitsui Chemicals, Nihon Neuroprotective vaccine -- University of Schering Auckland MSH fusion toxin - Ligand neurotrophic chimaeras - Regeneron MSI-99 -- Genaera neurotrophic factor -- NsGene, CereMedix MT 201 -- Micromet

#### **FIG. 28U**

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NeuroVax -- Immune Response Oncophage -- Antigenics Oncostatin M -- Bristol-Myers Squibb neurturin - Genentech OncoVax-CL -- Jenner Biotherapies neutral endopeptidase - Genentech OncoVax-P -- Jenner Biotherapies NGF enhancers -- NeuroSearch NHL vaccine -- Large Scale Biology onercept -- Yeda NIP45 -- Boston Life Sciences onychomycosis vaccine -- Boehringer Ingelheim NKI-B20 opebecan - XOMA NM 01 - Nissin Food opioids -- Arizona NMI-139 -- NitroMed Oprelvekin – Genetics Institute NMMP -- Genetics Institute Oregovomab -- AltaRex NN-2211 -- Novo Nordisk Org-33408 b- Akzo Nobel Noggin - Regeneron Orolip DP --- EpiCept Nonacog alfa oryzacystatin Norelin -- Biostar OSA peptides – GenSci Regeneration Norwalk virus vaccine osteoblast-cadherin GF -- Pharis NRLU 10 -- NeoRx Osteocalcin-thymidine kinase gene therapy NRLU 10 PE -- NeoRx osteogenic protein -- Curis NT-3 -- Regeneron osteopontin -- OraPharma NT-4/5 -- Genentech osteoporosis peptides -- Integra, Telios NU 3056 osteoprotegerin - Amgen, SnowBrand NU 3076 otitis media vaccines - Antex Biologics NX 1838 -- Gilead Sciences ovarian cancer -- University of Alabama NY ESO-1/CAG-3 antigen -- NIH OX40-lgG fusion protein -- Cantab, Xenova NYVAC-7 -- Aventis Pasteur P 246 -- Diatide NZ-1002 -- Novazyme P 30 -- Alfacell obesity therapy - Nobex p1025 - Active Biotech OC 10426 -- Ontogen P-113<sup>^</sup> - Demegen OC 144093 -- Ontogen P-16 peptide -- Transition Therapeutics OCIF -- Sankyo p43 -- Ramot Oct-43 -- Otsuka P-50 peptide -- Transition Therapeutics Odulimomab - Immunotech p53 + RAS vaccine -- NIH, NCI OK PSA - liposomal PACAP(1-27) analogue OKT3-gamma-1-ala-ala paediatric vaccines -- Chiron OM 991 Pafase - ICOS OM 992 PAGE-4 plasmid DNA -- IDEC Omalizumab -- Genentech PAI-2 -- Biotech Australia, Human oncoimmunin-L -- NIH Therapeutics. Oncolysin B -- ImmunoGen Palifermin (keratinocyte growth factor) --Oncolysin CD6 -- ImmunoGen Amaen Oncolysin M -- ImmunoGen Palivizumab -- MedImmune Oncolysin S - ImmunoGen

# **FIG. 28V**

PEG-uricase -- Mountain View PAM 4 -- Merck Pegvisomant -- Genentech pamiteplase -- Yamanouchi PEGylated proteins, PolyMASC -- Valentis pancreatin, Minitabs -- Eurand PEGylated recombinant native human leptin Pangen -- Fournier Pantarin - Selective Genetics -- Roche Parainfluenza virus vaccine - Pharmacia, Pemtumomab Penetratin -- Cyclacel Pierre Fabre Pepscan - Antisoma paraoxanase - Esperion parathyroid hormone - Abiogen, Korea peptide G - Peptech, ICRT peptide vaccine -- NIH, NCI Green Cross Pexelizumab Parathyroid hormone (1-34) -pexiganan acetate -- Genaera Chugai/Suntory Pharmaprojects No. 3179 -- NYU Parkinson's disease gene therapy -- Cell Pharmaprojects No. 3390 -- Ernest Orlando Genesys/ Ceregene Parvovirus vaccine -- MedImmune Pharmaprojects No. 3417 -- Sumitomo Pharmaprojects No. 3777 -- Acambis PCP-Scan - Immunomedics Pharmaprojects No. 4209 -- XOMA PDGF -- Chiron Pharmaprojects No. 4349 – Baxter Intl. PDGF cocktail -- Theratechnologies Pharmaprojects No. 4651 peanut allergy therapy -- Dynavax Pharmaprojects No. 4915 -- Avanir PEG anti-ICAM MAb -- Boehringer Pharmaprojects No. 5156 -- Rhizogenics Ingelheim Pharmaprojects No. 5200 -- Pfizer PEG asparaginase -- Enzon ' Pharmaprojects No. 5215 -- Origene PEG glucocerebrosidase Pharmaprojects No. 5216 -- Origene PEG hirudin -- Knoll Pharmaprojects No. 5218 - Origene PEG interferon-alpha-2a -- Roche Pharmaprojects No. 5267 -- ML PEG interferon-alpha-2b + ribavirin -Laboratories Biogen, Enzon, ICN Pharmaceuticals, Pharmaprojects No. 5373 -- MorphoSys Schering-Plough Pharmaprojects No. 5493 -- Metabolex PEG MAb A5B7 -Pharmaprojects No. 5707 -- Genentech Pegacaristim - Amgen -- Kirin Brewery --Pharmaprojects No. 5728 -- Autogen ZymoGenetics Pharmaprojects No. 5733 -- BioMarin Pegaldesleukin -- Research Corp Pharmaprojects No. 5757 -- NIH pegaspargase -- Enzon Pharmaprojects No. 5765 -- Gryphon pegfilgrastim -- Amgen Pharmaprojects No. 5830 -- AntiCancer PEG-interferon Alpha -- Viragen Pharmaprojects No. 5839 -- Dyax PEG-interferon Alpha 2A -- Hoffman La-Pharmaprojects No. 5849 -- Johnson & Roche Johnson PEG-interferon Alpha 2B -- Schering-Pharmaprojects No. 5860 -- Mitsubishi-Plough Tokyo PEG-r-hirudin -- Abbott PEG-rHuMGDF -- Amgen

#### **FIG. 28W**

Plasminogen activators -- Abbott Pharmaprojects No. 5869 -- Oxford Laboratories, American Home Products, GlycoSciences | Boehringer Mannheim, Chiron Pharmaprojects No. 5883 -- Asahi Brewery Pharmaprojects No. 5947 - StressGen Corporation, DuPont Pharmaceuticals, Eli Lilly, Shionogi, Genentech, Genetics Pharmaprojects No. 5961 --Institute, GlaxoSmithKline, Hemispherx Theratechnologies Pharmaprojects No. 5962 - NIH Biopharma, Merck & Co, Novartis, Pharmacia Corporation, Wakamoto, Yeda Pharmaprojects No. 5966 -- NIH plasminogen-related peptides -- Bio-Tech. Pharmaprojects No. 5994 -- Pharming Pharmaprojects No. 5995 -- Pharming General/MGH Pharmaprojects No. 6023 - IMMUCON platelet factor 4 -- RepliGen Platelet-derived growth factor - Amgen --Pharmaprojects No. 6063 - Cytoclonal ZymoGenetics Pharmaprojects No. 6073 -- SIDDCO plusonermin-- Hayashibara Pharmaprojects No. 6115 -- Genzyme PMD-2850 -- Protherics Pharmaprojects No. 6227 - NIH Pneumococcal vaccine -- Antex Biologics, Pharmaprojects No. 6230 -- NIH Pharmaprojects No. 6236 -- NIH Aventis Pasteur Pneumococcal vaccine intranasal --Pharmaprojects No. 6243 -- NIH BioChem Vaccines/Biovector Pharmaprojects No. 6244 -- NIH PR1A3 Pharmaprojects No. 6281 - Senetek PR-39 Pharmaprojects No. 6365 - NIH pralmorelin -- Kaken Pharmaprojects No. 6368 - NIH Pretarget-Lymphoma -- NeoRx Pharmaprojects No. 6373 -- NIH Pharmaprojects No. 6408 -- Pan Pacific Priliximab -- Centocor PRO 140 -- Progenics Pharmaprojects No. 6410 – Athersys PRO 2000 -- Procept Pharmaprojects No. 6421 -- Oxford PRO 367 -- Progenics **GlycoSciences** PRO 542 -- Progenics Pharmaprojects No. 6522 -- Maxygen pro-Apo A-I -- Esperion Pharmaprojects No. 6523 -- Pharis prolactin -- Genzyme Pharmaprojects No. 6538 -- Maxygen Prosaptide TX14(A) -- Bio-Tech. General Pharmaprojects No. 6554 -- APALEXO prostate cancer antbodies -- Immunex, Pharmaprojects No. 6560 -- Ardana Pharmaprojects No. 6562 -- Bayer **UroCor** prostate cancer antibody therapy --Pharmaprojects No. 6569 -- Eos Genentech/UroGenesys. Phenoxazine Genotherapeutics Phenylase -- Ibex prostate cancer immunotherapeutics -- The Pigment epithelium derived factor --**PSMA Development Company** plasminogen activator inhibitor-1. recombinant -- DuPont Pharmaceuticals - prostate cancer vaccine -- Aventis Pasteur, Zonagen, Corixa, Dendreon, Jenner Biotherapies, Therion Biologics

FIG. 28X

RD 62198 prostate-specific antigen -- EntreMed rDnase -- Genentech protein A -- RepliGen RDP-58 -- SangStat protein adhesives -- Enzon protein C -- Baxter Intl., PPL Therapeutics, RecepTox-Fce - Keryx RecepTox-GnRH - Keryx, MTR ZymoGenetics **Technologies** protein C activator - Gilead Sciences RecepTox-MBP - Keryx, MTR protein kinase R antags -- NIH **Technologies** protirelin - Takeda protocadherin 2 -- Caprion recFSH -- Akzo Nobel, Organon REGA 3G12 Pro-urokinase - Abbott, Bristol-Myers Squibb, Dainippon, Tosoh -- Welfide Regavirumab -- Teilin P-selectin glycoprotein ligand-1 -- Genetics relaxin -- Connetics Corp Renal cancer vaccine -- Macropharm Institute pseudomonal infections -- InterMune repifermin -- Human Genome Sciences Respiratory syncytial virus PFP-2 vaccine --Pseudomonas vaccine - Cytovax Wyeth-Lederle PSGL-Ig -- American Home Products Respiratory syncytial virus vaccine --PSP-94 -- Procyon GlaxoSmithKline, Pharmacia, Pierre Fabre PTH 1-34 -- Nobex Respiratory syncytial virus vaccine Quilimmune-M -- Antigenics inactivated R 744 -- Roche Respiratory syncytial virus-parainfluenza R 101933 virus vaccine -- Aventis Pasteur, R 125224 -- Sankyo Pharmacia RA therapy -- Cardion Reteplase -- Boehringer Mannheim, Rabies vaccine recombinant -- Aventis Hoffman La-Roche Pasteur, BioChem Vaccines, Kaketsuken Retropep -- Retroscreen Pharmaceuticals | RadioTheraClM - YM BioSciences RFB4 (dsFv) PE38 RFI 641 – American Home Products Ramot project No. 1315 -- Ramot RFTS - UAB Research Foundation Ramot project No. K-734A - Ramot Ramot project No. K-734B -- Ramot RG 12986 -- Aventis Pasteur RG 83852 -- Aventis Pasteur Ranibizumab (Anti-VEGF fragment) --RG-1059 - RepliGen Genentech rGCR -- NIH RANK -- Immunex ranpirnase -- Alfacell rGLP-1 -- Restoragen ranpirnase-anti-CD22 MAb -- Alfacell rGRF -- Restoragen rh Insulin - Eli Lilly RANTES inhibitor — Milan RHAMM targeting peptides - Cangene RAPID drug delivery systems - ARIAD rHb1.1 - Baxter Intl. rasburicase -- Sanofi rBPI-21, topical -- XOMA rhCC10 - Claragen RC 529 -- Corixa rhCG - Serono · Rheumatoid arthritis gene therapy rCFTR - Genzyme Transgenics

#### **FIG. 28Y**

SB RA 31012 -Rheumatoid arthritis vaccine -- Veterans SC 56929 - Pharmacia Affairs Medical Center SCA binding proteins -- Curis, Enzon rhLH -- Serono scFv(14E1)-ETA Berlex Laboratories, Ribozyme gene therapy -- Genset Schering AG Rickettsial vaccine recombinant ScFv(FRP5)-ETA -RIGScan CR -- Neoprobe ScFv6C6-PE40 -RIP-3 -- Rigel SCH 55700 - Celltech Rituximab -- Genentech Schistosomiasis vaccine -- Glaxo RK-0202 -- RxKinetix Wellcome/Medeva, Brazil RLT peptide -- Esperion SCPF -- Advanced Tissue Sciences rM/NEI -- IVAX scuPA-suPAR complex -- Hadasit rmCRP -- Immtech SD-9427 -- Pharmacia RN-1001 -- Renovo SDF-1 -- Ono RN-3 -- Renovo SDZ 215918 - Novartis RNAse conjugate -- Immunomedics SDZ 280125 - Novartis RO 631908 -- Roche SDZ 89104 -- Novartis Rotavirus vaccine -- Merck SDZ ABL 364 -- Novartis RP 431 -- DuPont Pharmaceuticals SDZ MMA 383 -- Novartis RP-128 -- Resolution Secretin - Ferring, Repligen RPE65 gene therapy -serine protease inhibs -- Pharis RPR 110173 -- Aventis Pasteur sermorelin acetate -- Serono RPR 115135 -- Aventis Pasteur SERP-1 -- Viron RPR 116258A -- Aventis Pasteur sertenef - Dainippon rPSGL-Ig -- American Home Products serum albumin, Recombinant human r-SPC surfactant -- Byk Gulden Aventis Behring RSV antibody -- Medimmune serum-derived factor -- Hadasit Ruplizumab -- Biogen: Sevirumab -- Novartis rV-HER-2/neu -- Therion Biologics SGN 14 - Seatle Genetics SA 1042 -- Sankyo SGN 15 - Seatle Genetics sacrosidase -- Orphan Medical SGN 17/19 - Seatle Genetics Sant 7 SGN 30 - Seatle Genetics Sargramostim -- Immunex SGN-10 -- Seatle Genetics saruplase -- Gruenenthal SGN-11 -- Seatle Genetics Satumomab -- Cytogen SH 306 - DuPont Pharmaceuticals SB 1 - COR Therapeutics Shanvac-B -- Shantha SB 207448 - GlaxoSmithKline Shigella flexneri vaccine - Avant, Acambis, SB 208651 — GlaxoSmithKline Novavax SB 240683 -- GlaxoSmithKline Shigella sonnei vaccine -SB 249415 -- GlaxoSmithKline sICAM-1 -- Boehringer Ingelheim SB 249417 -- GlaxoSmithKline Silteplase -- Genzyme SB 6 - COR Therapeutics

FIG. 28Z

Staphylococcus aureus vaccine conjugate --SIV vaccine -- Endocon, Institut Pasteur SK 896 -- Sanwa Kagaku Kenkyusho Nabi Staphylococcus therapy - Tripep SK-827 -- Sanwa Kagaku Kenkyusho Staphylokinase - Biovation, Prothera, Skeletex -- CellFactors Thrombogenetics SKF 106160 - GlaxoSmithKline Streptococcal A vaccine -- M6 S-nitroso-AR545C --Pharmaceuticals, North American Vaccine SNTP - Active Biotech Streptococcal B vaccine -- Microscience somatomedin-1 - GroPep, Mitsubishi-Streptococcal B vaccine recombinant --Tokyo, NIH **Biochem Vaccines** somatomedin-1 carrier protein -- Insmed Streptococcus pyogenes vaccine somatostatin -- Ferring STRL-33 -- NIH Somatotropin/ Subalin -- SRC VB VECTOR Human Growth Hormone -- Bio-Tech. SUIS -- United Biomedical General, Eli Lilly somatropin -- Bio-Tech. General, Alkermes, SUIS-LHRH - United Biomedical ProLease, Aventis Behring, Biovector, SUN-E3001 -- Suntory super high affinity monoclonal antibodies -Cangene, Dong-A, Eli Lilly, Emisphere, Enact, Genentech, Genzyme Transgenics, YM BioSciences Grandis/InfiMed, CSL, InfiMed, MacroMed, Superoxide dismutase - Chiron, Enzon, Ube Industries, Bio-Tech, Yeda Novartis, Novo Nordisk, Pharmacia superoxide dismutase-2 -- OXIS Serono, TranXenoGen suppressin - UAB Research Foundation somatropin derivative - Schering AG SY-161-P5 - ThromboGenics somatropin, AIR - Eli Lilly SY-162 -- ThromboGenics Somatropin, inhaled - Eli Lilly/Alkermes Systemic lupus erythematosus vaccine -somatropin, Kabi - Pharmacia MedClone/VivoRx somatropin, Orasome -- Novo Nordisk T cell receptor peptides -- Xoma Sonermin - Dainippon Pharmaceutical SP(V5.2)C - Supertek T cell receptor peptide vaccine T4N5 liposomes -- AGI Dermatics SPf66 TACI, soluble -- ZymoGenetics sphingomyelinase - Genzyme targeted apoptosis -- Antisoma SR 29001 - Sanofi tasonermin -- Boehringer Ingelheim SR 41476 -- Sanofi SR-29001 - Sanofi TASP TASP-V SS1(dsFV)-PE38 - NeoPharm Tat peptide analogues -- NIH ß2 microglobulin -- Avidex TBP I - Yeda ß2-microglobulin fusion proteins -- NIH TBP II **ß-amyloid peptides -- CeNeS** TBV25H - NIH ß-defensin -- Pharis Tc 99m ior cea1 - Center of Molecular Staphylococcus aureus infections -**Immunology** Inhibitex/ZLB Tc 99m P 748 -- Diatide

#### FIG. 28AA

TIF -- Xoma

Tifacogin -- Chiron, NIS, Pharmacia

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Tissue factor - Genentech Tc 99m votumumab - Intracell Tc-99m rh-Annexin V - Theseus Imaging Tissue factor pathway inhibitor TJN-135 -- Tsumura teceleukin -- Biogen TM 27 - Avant tenecteplase - Genentech Teriparatide - Armour Pharmaceuticals, TM 29 - Avant TMC-151 - Tanabe Seiyaku Asahi Kasei, Eli Lilly TNF tumour necrosis factor -- Asahi Kasei terlipressin -- Ferring TNF Alpha -- Cytlmmune testisin -- AMRAD TNF antibody -- Johnson & Johnson Tetrafibricin -- Roche TNF binding protein - Amgen TFPI -- EntreMed TNF degradation product - Oncotech toD-IL-2 - Takeda TNF receptor - Immunex TGF-Alpha -- ZymoGenetics TNF receptor 1, soluble -- Amgen TGF-ß -- Kolon TNF Tumour necrosis factor-alpha -- Asahi TGF-B2 -- Insmed Kasei, Genetech, Mochida TGF-ß3 -- OSI Thalassaemia gene therapy -- Crucell TNF-Alpha inhibitor -- Tripep TNFR:Fc gene therapy - Targeted Genetics TheraClM-h-R3 -- Center of Molecular Immunology, YM BioSciences TNF-SAM2 ToleriMab - Innogenetics Theradigm-HBV -- Epimmune Toxoplasma gondii vaccine --Theradigm-HPV -- Epimmune GlaxoSmithKline Theradigm-malaria -- Epimmune TP 9201 -- Telios Theradigm-melanoma -- Epimmune TP10 - Avant TheraFab - Antisoma ThGRF 1-29 - Theratechnologies TP20 -- Avant ThGRF 1-44 -- Theratechnologies tPA -- Centocor trafermin -- Scios Thrombin receptor activating peptide --TRAIL/Apo2L -- Immunex Abbott TRAIL-R1 MAb - Cambridge Antibody thrombomodulin - Iowa, Novocastra Technologies Thrombopoietin - Dragon Pharmaceuticals, transferrin-binding proteins - CAMR Genentech Transforming growth factor-beta-1 -thrombopoietin, Pliva -- Receptron Genentech Thrombospondin 2 transport protein -- Genesis thrombostatin - Thromgen Trastuzumab - Genetech thymalfasin - SciClone TRH -- Ferring thymocartin - Gedeon Richter thymosin Alpha1 - NIH Triabin -- Schering AG Triconal · thyroid stimulating hormone -- Genzyme Triflavin tICAM-1 -- Bayer troponin I - Boston Life Sciences Tick anticoagulant peptide - Merck

#### FIG. 28BB

TRP-2<sup>^</sup> -- NIH

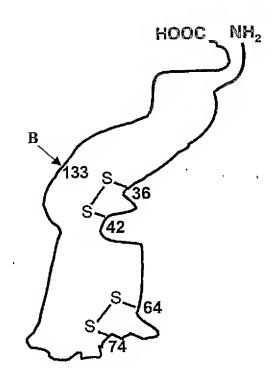
trypsin inhibitor -- Mochida

Vascular endothelial growth factors - R&D TSP-1 gene therapy -TT-232 vascular targeting agents -- Peregrine TTS-CD2 -- Active Biotech vasopermeation enhancement agents --Tuberculosis vaccine - Aventis Pasteur, Peregrine Genesis vasostatin - NIH Tumor Targeted Superantigens - Active VCL - Bio-Tech, General Biotech -- Pharmacia VEGF - Genentech, Scios tumour vaccines -- PhotoCure tumour-activated prodrug antibody VEGF inhibitor -- Chugai VEGF-2 -- Human Genome Sciences conjugates - Millennium/ImmunoGen VEGF-Trap -- Regeneron tumstatin - ILEX viscumin, recombinant - Madaus Tuvirumab -- Novartis Vitaxin TV-4710 – Teva Vitrase -- ISTA Pharmaceuticals TWEAK receptor -- Immunex West Nile virus vaccine -- Bavarian Nordic TXU-PAP WP 652 TY-10721 – TOA Eiyo Type I diabetes vaccine -- Research Corp WT1 vaccine -- Corixa WX-293 - Wilex BioTech. Typhoid vaccine CVD 908 WX-360 -- Wilex BioTech. U 143677 -- Pharmacia WX-UK1 -- Wilex BioTech. U 81749 -- Pharmacia XMP-500 -- XOMA UA 1248 – Arizona XomaZyme-791 -- XOMA UGIF - Sheffield XTL 001 - XTL Biopharmaceuticals UIC 2 XTL 002 -- XTL Biopharmaceuticals UK 101 yeast delivery system -- Globelmmune UK-279276 -- Corvas Intl. Yersinia pestis vaccine urodilatin - Pharis YIGSR-Stealth - Johnson & Johnson urofollitrophin - Serono Yissum Project No. D-0460 -- Yissum Urokinase -- Abbott YM 207 - Yamanouchi uteroferrin-- Pepgen YM 337 -- Protein Design Labs V 20 -- GLYCODesign Yttrium-90 labelled biotin V2 vasopressin receptor gene therapy Yttrium-90-labeled anti-CEA MAb T84.66 vaccines -- Active Biotech ZD 0490 - AstraZeneca Varicella zoster glycoprotein vaccine -ziconotide -- Elan Research Corporation Technologies Varicella zoster virus vaccine live -- Cantab ZK 157138 -- Berlex Laboratories Zolimomab aritox Pharmaceuticals Zorcell - Immune Response Vascular endothelial growth factor -

FIG. 28CC

Genentech, University of California

ZRXL peptides -- Novartis



$$\mathbf{B} \leftarrow \begin{pmatrix} (\operatorname{Sia})_b \\ -\operatorname{GalNAc-(Gal)}_a - (\operatorname{Sia})_c - (R)_d \end{pmatrix}_e$$

a-c, e (independently selected) = 0 or 1; d = 0; R = modifying group, sialyl or oligosialyl

FIG. 29A

CHO, BHK, 293 cells, Vero expressed G-CSF a-c, e (independently selected) = 0 or 1; d = 0

- 1. Sialidase
  - 2. CMP-SA-PEG, ST3Gal1

a-d, e (independently selected) = 0 or 1; R = PEG.

# FIG. 29B

Insect cell expressed G-CSF a, e (independently selected) = 0 or 1; b, c, d = 0.

- 1. Galactosyltransferase, UDP-Gal
  2. CMP-SA-PEG, ST3Gal1
- a, c, d, e (independently selected) = 0 or 1; R = PEG.

FIG. 29C

```
E. coli expressed G-CSF a-e=0.
```

- 1. GalNAc Transferase, UDP-GalNAc
- ↓ 2. CMP-SA-PEG, sialyltransferase

```
c, d, e (independently selected) = 0 or 1;
a, b = 0; R = PEG.
```

# FIG. 29D

NSO expressed G-CSF a, e (independently selected) = 0 or 1; b, c, d = 0

> 1. CMP-SA-levulinate, ST3Gal1 2. H<sub>4</sub>N<sub>2</sub>-PEG

a, c, d, e (independently selected) = 0 or 1; b = 0; R = PEG.

# FIG. 29E

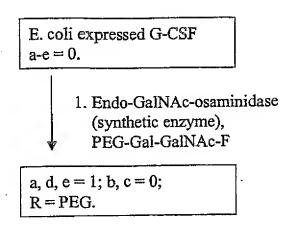


FIG. 29F

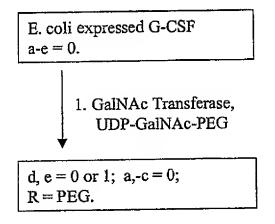
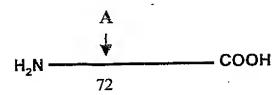


FIG. 29G



$$A \leftarrow \begin{array}{c} \text{(Fuc)}_{i} \\ \text{-GlcNAc-Man} \\ \text{([GlcNAc-(Gal)_{a}]_{e}-(Sia)_{j}-(R)_{v})_{r}} \\ \text{([GlcNAc-(Gal)_{b}]_{f}-(Sia)_{k}-(R)_{w})_{s}} \\ \text{(R')}_{dd} \\ \text{([GlcNAc-(Gal)_{d}]_{h}-(Sia)_{m}-(R)_{y})_{u}} \\ \text{([GlcNAc-(Gal)_{d}]_{h}-(Sia)_{m}-(R)_{y}} \\ \text{([GlcNAc-(Gal)_{d}]_{h}-(Sia)_{m}$$

a-d, i, n-u (independently selected) = 0 or 1.

aa, bb, cc, dd, ee (independently selected) = 0 or 1.

e-h (independently selected) = 0 to 6.

j-m (independently selected) = 0 to 20.

v-z = 0; R = modifying group, mannose, oligo-mannose.

R' = H, glycosyl residue, modifying group,
glycoconjugate.

FIG. 30A

```
ŒIO, BHK, 293 cells, Vero expressed interferon alpha 14C.
a-d, aa, bb = 1; e-h = 1 to 4;
cc, j-m, i, r-u (independently selected) = 0 or 1;
q, n-p, v-z, cc, dd, ee = 0.
```

- 1. Sialidase
- 2. CMP-SA-PEG, ST3Gal3

```
a-d, aa, bb = 1; e-h = 1 to 4;
bb, cc, i, r-u (independently selected) = 0 or 1;
q, n-p, v-z, cc, dd, ee = 0;
v-y (independently selected) = 1,
when j-m (independently selected) = 1;
R = PEG.
```

# **FIG. 30B**

```
Insect cell or fungi expressed interferon alpha-14C. a-d, f, h, j-q, s, u, v-z, cc, dd, ee = 0; e, g, i, r, t (independently selected) = 0 or 1; aa, bb = 1.
```

- GNT's 1&2, UDP-GlcNAc
   Galactosyltransferase, UDP-Gal-PEG
- b, d, f, h, j-q, s, u, w, y, z, cc, dd, ee = 0; a, c, e, g, i, r, t, v, x (independently selected) = 0 or 1; v, x (independently selected) = 1, when a, c, (independently selected) = 1; aa, bb = 1; R = PEG.

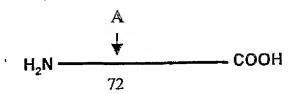
## FIG. 30C

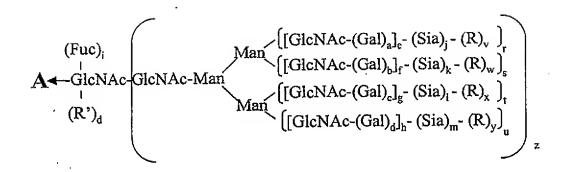
Yeast expressed interferon alpha-14C. a-q, cc, dd, ee, v-z = 0; r-y (independently selected) = 0 to 1; aa, bb = 1;R (branched or linear) = Man, oligomannose or polysaccharide.

- 1. Endo-H
- 2. Galactosyltransferase, UDP-Gal 3.. CMP-SA-PEG, ST3Gal3

$$a-z$$
,  $bb=0$ ;  $aa=1$ ;  $R'=-Gal-Sia-PEG$ .

FIG. 30D





a-d, i, r-u (independently selected) = 0 or 1. e-h (independently selected) = 0 to 4. j-m (independently selected) = 0 or 1. n, v-y = 0; z=0 or 1. R = polymer; R' = sugar, glycoconjugate.

FIG. 30E

```
CHO, BHK, 293 cells, Vero expressed interferon alpha-14C.

h = 1 to 3;
a-g, j-m, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
n, v-y = 0; z = 1.
```

## 1. CMP-SA-PEG, ST3Gal3

```
h = 1 to 3;
a-g, i (independently selected) = 0 or 1;
r-u (independently selected) = 0 or 1;
j-m, v-y (independently selected) = 0 or 1;
z = 1; n = 0; R = PEG.
```

# FIG. 30F

```
Insect cell or fungi expressed interferon alpha-14C.

a-d, f, h, j-n, s, u, v-y = 0;

e, g, i, r, t (independently selected) = 0 or 1;

z = 1.
```

- 1. GNT's 1,2,4,5, UDP-GlcNAc
- 2. Galactosyltransferase, UDP-Gal
- 3. CMP-SA-PEG, ST3Gal3

a-m, r-y (independently selected) = 0 or 1; z = 1; n = 0; R = PEG.

## FIG. 30G

Yeast expressed interferon alpha-14C. a-n = 0; r-y (independently selected) = 0 to 1; z = 1; R (branched or linear) = Man, oligomannose.

- 1. mannosidases
- 2. GNT's 1,2,4,5, UDP-GlcNAc
- 3. Galactosyltransferase, UDP-Gal
- 4.. CMP-SA-PEG, ST3Gal3

a-m, r-y (independently selected) = 0 or 1; z = 1; n = 0; R = PEG.

## FIG. 30H

NSO expressed interferon alpha 14C. a-i, r-u (independently selected) = 0 or 1; j-m, n, v-y = 0; z = 1.

1. CMP-SA-levulinate, ST3Gal3, buffer, salt
2. H<sub>4</sub>N<sub>2</sub>-PEG

a-i, j-m, r-y (independently selected) = 0 or 1; n = 0; z = 1; R = PEG.

FIG. 301